

**UTILITY PATENT APPLICATION TRANSMITTAL
(Large Entity)***(Only for new nonprovisional applications under 37 CFR 1.53(b))*Docket No.
A0000180-66-BDTotal Pages in this Submission
66**TO THE ASSISTANT COMMISSIONER FOR PATENTS**Box Patent Application
Washington, D.C. 20231

Transmitted herewith for filing under 35 U.S.C. 111(a) and 37 C.F.R. 1.53(b) is a new utility patent application for an invention entitled:

**SECRETED SOLUBLE ALPHA2DELTA-2, ALPHA2DELTA-3 OR ALPHA2DELTA-4 CALCIUM CHANNEL
SUBUNIT POLYPEPTIDES AND SCREENING ASSAYS USING SAME**

and invented by:

FRANCOIS BERTELLI, JASON BROWN, VISAKA DISSANAYAKEIf a **CONTINUATION APPLICATION**, check appropriate box and supply the requisite information:☐ Continuation ☐ Divisional ☐ Continuation-in-part (CIP) of prior application No.: _____

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Enclosed are:

Application Elements

1. ☒ Filing fee as calculated and transmitted as described below
2. ☒ Specification having 62 pages and including the following:
 - a. ☒ Descriptive Title of the Invention
 - b. ☐ Cross References to Related Applications *(if applicable)*
 - c. ☐ Statement Regarding Federally-sponsored Research/Development *(if applicable)*
 - d. ☐ Reference to Microfiche Appendix *(if applicable)*
 - e. ☒ Background of the Invention
 - f. ☒ Brief Summary of the Invention
 - g. ☐ Brief Description of the Drawings *(if drawings filed)*
 - h. ☒ Detailed Description
 - i. ☒ Claim(s) as Classified Below
 - j. ☒ Abstract of the Disclosure

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Application Elements (Continued)

3. ☐ Drawing(s) *(when necessary as prescribed by 35 USC 113)*
- a. ☐ Formal Number of Sheets _____
- b. ☐ Informal Number of Sheets _____
4. ☐ Oath or Declaration
- a. ☐ Newly executed *(original or copy)* ☐ Unexecuted
- b. ☐ Copy from a prior application (37 CFR 1.63(d)) *(for continuation/divisional application only)*
- c. ☐ With Power of Attorney ☐ Without Power of Attorney
- d. ☐ DELETION OF INVENTOR(S)
Signed statement attached deleting inventor(s) named in the prior application,
see 37 C.F.R. 1.63(d)(2) and 1.33(b).
5. ☐ Incorporation By Reference *(usable if Box 4b is checked)*
The entire disclosure of the prior application, from which a copy of the oath or declaration is supplied
under Box 4b, is considered as being part of the disclosure of the accompanying application and is hereby
incorporated by reference therein.
6. ☐ Computer Program in Microfiche *(Appendix)*
7. ☐ Nucleotide and/or Amino Acid Sequence Submission *(if applicable, all must be included)*
- a. ☐ Paper Copy
- b. ☐ Computer Readable Copy *(identical to computer copy)*
- c. ☐ Statement Verifying Identical Paper and Computer Readable Copy

Accompanying Application Parts

8. ☐ Assignment Papers *(cover sheet & document(s))*
9. ☐ 37 CFR 3.73(B) Statement *(when there is an assignee)*
10. ☐ English Translation Document *(if applicable)*
11. ☐ Information Disclosure Statement/PTO-1449 ☐ Copies of IDS Citations
12. ☐ Preliminary Amendment
13. ☒ Acknowledgment postcard
14. ☒ Certificate of Mailing
- ☐ First Class ☒ Express Mail *(Specify Label No.):* EJ881683366US

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Accompanying Application Parts (Continued)

15. ☐ Certified Copy of Priority Document(s) *(if foreign priority is claimed)*

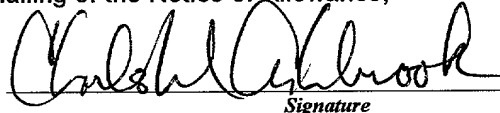
16. ☐ Additional Enclosures *(please identify below):*

Fee Calculation and Transmittal

CLAIMS AS FILED

For	#Filed	#Allowed	#Extra	Rate	Fee
Total Claims	22	- 20 =	2	x \$18.00	\$36.00
Indep. Claims	10	- 3 =	7	x \$78.00	\$546.00
Multiple Dependent Claims (check if applicable) <input type="checkbox"/>					\$0.00
BASIC FEE					\$760.00
OTHER FEE (specify purpose)					
TOTAL FILING FEE					\$1,342.00

- ☐ A check in the amount of _____ to cover the filing fee is enclosed.
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- ☐ Charge the issue fee set in 37 C.F.R. 1.18 at the mailing of the Notice of Allowance, pursuant to 37 C.F.R. 1.311(b).


Signature

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CERTIFICATE OF MAILING BY "EXPRESS MAIL" (37 CFR 1.10)Applicant(s): **Francois Bertelli, et al.**

Docket No.

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Serial No.

Filing Date

Examiner

Group Art Unit

Invention: **SECRETED SOLUBLE ALPH2DELTA-2, ALPHA2DELTA-3 OR ALPHA2DELTA-4 CALCIUM CHANNEL SUBUNIT POLYPEPTIDES AND SCREENING ASSAYS USING SAME**

I hereby certify that this Utility Patent Application Under 37 CFR 1.53(b)
(Identify type of correspondence)
is being deposited with the United States Postal Service "Express Mail Post Office to Addressee" service under
37 CFR 1.10 in an envelope addressed to: The Assistant Commissioner for Patents, Washington, D.C. 20231
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TITLE OF THE INVENTION: Secreted soluble $\alpha_2\delta$ -2, $\alpha_2\delta$ -3 or $\alpha_2\delta$ -4 calcium channel subunit polypeptides and screening assays using same

5 BACKGROUND OF THE INVENTION

Voltage-dependent Ca^{2+} channels (VDCCs) are heteromultimeric complexes present in both neuronal and non-neuronal tissues, including heart and skeletal muscle. VDCCs are minimally composed of three subunits: a pore-forming transmembrane α_1 subunit, a hydrophilic intracellular β subunit, and a membrane-associated $\alpha_2\delta$ subunit; a transmembrane γ subunit is also found in skeletal muscle tissue. Multiple subtypes and/or splice variants of the α_1 , β , and $\alpha_2\delta$ subunits have been found.

Gabapentin ((1-aminomethyl)cyclohexane acetic acid or Neurontin) is a structural analogue of GABA, which is mainly used as an adjunctive therapy for epilepsy. Recent research suggests that gabapentin may also have clinical utility for various indications including anxiety and pain. Although designed as a lipophilic GABA-mimetic, gabapentin does not have a high affinity for either GABA_A or GABA_B receptors, GABA uptake sites, or the GABA-degrading enzyme GABA-transaminase (EC 2.6.1.19).

A novel high affinity binding site for [^3H]gabapentin in rat, mouse, and porcine brains has been characterized. Recently, the [^3H]gabapentin-binding protein was isolated from pig brain and identified as the $\alpha_2\delta$ -1 subunit of VDCCs. None of the prototypic anticonvulsant drugs displace [^3H]gabapentin binding from the $\alpha_2\delta$ -1 subunit. [^3H]Gabapentin-binding is stereospecifically inhibited by two enantiomers of 3-isobutyl GABA. The rank order of potency of gabapentin, and S- and R-isobutyl GABA, at the [^3H]gabapentin binding site mirrors their anticonvulsant activity in mice. However, electrophysiological studies have yielded conflicting data on the action of gabapentin at VDCCs.

The $\alpha_2\delta$ subunit is derived from a single gene, the product of which is extensively post-translationally modified particularly through the cleavage of the signal sequence. The polypeptide is cleaved to form disulfide-bridged α_2 and δ peptides, both of which are heavily glycosylated. Although it seems clear today that the α_2 and δ peptides are membrane-associated peptides, it is unclear whether these peptides comprise one or several transmembrane domains. Furthermore, the location, size and structural configuration of these eventual transmembrane domains remains to be determined.

But in any event, the fact that $\alpha_2\delta$ is a membrane-associated protein, regardless of its precise structural configuration, renders its large scale expression in recombinant systems difficult. Indeed, as the $\alpha_2\delta$ protein is targeted to the membrane, it requires detergent solubilisation to release it for purification. This important drawback imposes considerable

5 restrictions for any potential applications requiring large amounts of recombinant protein. Furthermore, the various subtypes of $\alpha_2\delta$ subunits are different proteins with very low homologies. It is therefore extremely difficult to predict their respective behaviors, for example in gene truncation experiments.

10 The only assay currently available for the screening of ligands that bind the $\alpha_2\delta$ subunit involves the use of pig membrane extracts as a source of the $\alpha_2\delta$ subunit. Such an assay presents major inconvenients. Firstly, because the assay material is a membrane extract, it is very difficult to accurately determine the protein composition from one assay preparation to another particularly with regard to the subtype. Also, the presence of various impurities

15 in the assay preparation is a problem in small plate assays. Furthermore, as the protein preparation lacks homogeneity, the interaction between the targeted protein and the assay plate is often quite uneven. This rendres the streamlining of the assay in a high throughput format almost impossible to achieve.

20 SUMMARY OF THE INVENTION

In the context of the present invention, the inventors have found that it was possible to delete a portion of the nucleotide sequence encoding a eukaryotic, preferably a mammal

25 cerebral cortical voltage-dependent calcium channel α_2 subunit to yield a soluble secreted protein which retains its affinity for [^3H]gabapentin.

The invention concerns:

- 30 1) A purified or isolated nucleic acid encoding a mammalian secreted soluble cerebral cortical voltage-dependent calcium channel $\alpha_2\delta$ -2, $\alpha_2\delta$ -3 or $\alpha_2\delta$ -4 subunit polypeptide.
- 2) A purified or isolated nucleic acid according to 1), comprising a polynucleotide having at least 90% identity with the sequence encoding :

- from amino-acid 1 to between amino-acids 1027 and 1062 of SEQ ID N°20 for $\alpha_2\delta$ -2,
- from amino-acid 1 to between amino-acids 984 and 1019 of SEQ ID N°22 for $\alpha_2\delta$ -3.

3) A purified or isolated nucleic acid according to 1), having at least 90% identity with the sequence encoding :

- from amino-acid 1 to between amino-acids 1047 and 1062 of SEQ ID N°20 for $\alpha_2\delta$ -2,
- from amino-acid 1 to between amino-acids 1004 and 1019 of SEQ ID N°22 for $\alpha_2\delta$ -3.

4) A purified or isolated nucleotide sequence according to 1) wherein said sequence is the sequence of SEQ ID N°1, SEQ ID N°2, SEQ ID N°3, SEQ ID N°7, SEQ ID N°8, SEQ ID N°9, SEQ ID N°13, SEQ ID N°14, SEQ ID N°15, SEQ ID N°19 or SEQ ID N°21.

5) A purified or isolated nucleic acid, having at least 90% identity with the nucleotide sequence of SEQ ID N°19 or SEQ ID N°21.

6) A purified or isolated polynucleotide comprising at least 10 consecutive nucleotides of the nucleotide sequence of SEQ ID N°19 or SEQ ID N°21.

7) A polynucleotide probe or primer hybridizing, under stringent conditions, with the with the nucleotide sequence of SEQ ID N°19 or SEQ ID N°21.

8) A method for the amplification of a nucleic acid encoding a mammalian secreted soluble cerebral cortical voltage-dependent calcium channel $\alpha_2\delta$ -n subunit polypeptide wherein n is 2, 3 or 4, said method comprising the steps of:

- (a) contacting a test sample suspected of containing the target secreted soluble $\alpha_2\delta$ -n subunit nucleic acid, or a sequence complementary thereto, with an amplification reaction reagent comprising a pair of amplification primers located on either side of the $\alpha_2\delta$ -n subunit nucleic acid region to be amplified, and
- (b) optionally, detecting the amplification products.

9) A kit for the amplification of a nucleic acid encoding a secreted soluble $\alpha_2\delta$ -n subunit polypeptide wherein n is 2, 3 or 4, or a complementary sequence thereto in a test sample, wherein said kit comprises:

(a) a pair of oligonucleotide primers which can hybridize, under stringent conditions, to the secreted soluble $\alpha_2\delta$ -n subunit nucleic acid region to be amplified;

(b) optionally, the reagents necessary for performing the amplification reaction.

5

10) A recombinant vector comprising a nucleic acid according to 1).

11) A recombinant host cell comprising a nucleic acid according to 1).

10 12) A method for producing a secreted soluble $\alpha_2\delta$ -n subunit wherein n is 2, 3 or 4, and said method comprises the steps of:

(a) inserting the nucleic acid encoding the desired $\alpha_2\delta$ -n subunit polypeptide in an appropriate vector;

15

(b) culturing, in an appropriate culture medium, a host cell previously transformed or transfected with the recombinant vector of step (a);

(c) harvesting the culture medium thus obtained or lyse the host cell, for example by sonication or osmotic shock;

(d) separating or purifying, from said culture medium, or from the pellet of the resultant host cell lysate, the thus produced $\alpha_2\delta$ -n subunit polypeptide of interest.

20

13) A purified or isolated recombinant polypeptide comprising the amino acid sequence of a secreted soluble $\alpha_2\delta$ -2, $\alpha_2\delta$ -3 or $\alpha_2\delta$ -4 subunit polypeptide.

14) A recombinant polypeptide according to 14), having at least 80% amino-acid identity

25 with a polypeptide comprising :

- from amino acid 1 to between amino acids 1027 and 1062 of the amino acid sequence of SEQ ID N°20, or

- from amino acid 1 to between amino acids 1019 and 1079 of the amino acid sequence of SEQ ID N°22.

30

15) A recombinant polypeptide according to 14), wherein said recombinant polypeptide is selected from the group consisting of the amino acid sequences of SEQ ID n°4, SEQ ID n°5, SEQ ID n°6, SEQ ID n°10, SEQ ID n°11, SEQ ID n°12, SEQ ID n°16, SEQ ID n°17 and SEQ ID n°18.

16) A method for the screening of ligands which bind a cerebral cortical voltage-dependent calcium channel $\alpha_2\delta$ -n subunit wherein n is 2, 3 or 4, said method comprising the steps of:

- 5 - contacting a secreted soluble recombinant calcium channel $\alpha_2\delta$ -n subunit polypeptide with:
- a ligand of interest; and
 - a labelled compound which binds the $\alpha_2\delta$ -n subunit; and
- measuring the level of binding of the labelled compound to the $\alpha_2\delta$ -n subunit.

10

17) A method according to 16), wherein said method is a scintillation proximity assay.

18) A method according to 16), wherein said method is a flashplate assay.

15 19) A method according to 16), wherein said method is a filter binding assay.

20) A method according to 16), wherein said secreted soluble recombinant calcium channel $\alpha_2\delta$ -n subunit polypeptide is selected from polypeptides having at least 80%, preferably 90%, more preferably 95%, and most preferably 98 or 99% amino-acid identity with the polypeptide comprising from amino acid 1 to between amino-acids 984 and 1063, preferably between amino-acids 994 and 1054, and most preferably between amino-acids 1019 and 1054 of SEQ ID N°5 or SEQ ID N°16.

21) A method according to 16), wherein said secreted soluble recombinant calcium channel $\alpha_2\delta$ -n subunit polypeptide is selected from the group consisting of SEQ ID N°6, 7, 8, 9, 13, 14 and 15, with the polypeptides of SEQ ID N°9 and SEQ ID N°15 being most preferred.

22) A kit for the screening of ligands which bind a cerebral cortical voltage-dependent calcium channel $\alpha_2\delta$ -n subunit wherein n is 2, 3 or 4, said kit comprising:

- a secreted soluble recombinant calcium channel $\alpha_2\delta$ -n subunit; and
- a labelled compound which binds to the $\alpha_2\delta$ -n subunit.

Hence, the invention concerns nucleotide sequence fragments of a cerebral cortical voltage dependent calcium channel $\alpha_2\delta$ -2, $\alpha_2\delta$ -3 or $\alpha_2\delta$ -4 subunit cDNA encoding a soluble secreted $\alpha_2\delta$ -2, $\alpha_2\delta$ -3 or $\alpha_2\delta$ -4 subunit polypeptide (hereinafter a $\alpha_2\delta$ -2, $\alpha_2\delta$ -3 or $\alpha_2\delta$ -4 subunit). Preferably, these nucleotide sequences encode a soluble secreted $\alpha_2\delta$ -2, $\alpha_2\delta$ -3 or $\alpha_2\delta$ -4 subunit polypeptide bearing a gabapentin or a [3 H]gabapentin binding site. More preferably, the soluble secreted $\alpha_2\delta$ -2, $\alpha_2\delta$ -3 or $\alpha_2\delta$ -4 subunit nucleic acid is derived from a eukaryotic, preferably a mammal, more preferably a human $\alpha_2\delta$ -2, $\alpha_2\delta$ -3 or $\alpha_2\delta$ -4 subunit.

A further object of the present invention concerns recombinant vectors comprising a nucleic acid sequence encoding a soluble secreted $\alpha_2\delta$ -2, $\alpha_2\delta$ -3 or $\alpha_2\delta$ -4 subunit polypeptide.

The invention also encompasses host cells and transgenic non-human mammals comprising said nucleic acid sequences or recombinant vectors.

The invention also concerns a soluble secreted $\alpha_2\delta$ -2, $\alpha_2\delta$ -3 or $\alpha_2\delta$ -4 subunit polypeptide which is characterized in that it is a soluble secreted polypeptide having affinity for [3 H]gabapentin. Preferably, the soluble secreted polypeptide is derived from a mammal, more preferably a human $\alpha_2\delta$ -2, $\alpha_2\delta$ -3 or $\alpha_2\delta$ -4 subunit.

The inventors have also found that it was possible to use a soluble secreted form of a voltage-dependant calcium channel $\alpha_2\delta$ -2, $\alpha_2\delta$ -3 or $\alpha_2\delta$ -4 subunit polypeptide in an assay for the screening of ligands which bind the $\alpha_2\delta$ -2, $\alpha_2\delta$ -3 or $\alpha_2\delta$ -4 subunit.

The invention therefore also concerns a method for the screening of ligands which bind a calcium channel $\alpha_2\delta$ -2, $\alpha_2\delta$ -3 or $\alpha_2\delta$ -4 subunit.

The method comprises the steps of:

- contacting a secreted soluble recombinant calcium channel $\alpha_2\delta$ -2, $\alpha_2\delta$ -3 or $\alpha_2\delta$ -4 subunit polypeptide with:
 - a ligand of interest; and
 - a labelled compound which binds a $\alpha_2\delta$ -2, $\alpha_2\delta$ -3 or $\alpha_2\delta$ -4 subunit; and
- measuring the level of binding of the labelled compound to the secreted soluble $\alpha_2\delta$ -2, $\alpha_2\delta$ -3 or $\alpha_2\delta$ -4 subunit.

The invention also concerns a kit for the screening of ligands which bind a calcium channel $\alpha_2\delta$ -2, $\alpha_2\delta$ -3 or $\alpha_2\delta$ -4 subunit.

The kit comprises:

- a secreted soluble recombinant calcium channel $\alpha_2\delta$ -2, $\alpha_2\delta$ -3 or $\alpha_2\delta$ -4 subunit polypeptide; and
- a labelled compound which binds a calcium channel $\alpha_2\delta$ -2, $\alpha_2\delta$ -3 or $\alpha_2\delta$ -4 subunit.

10 DETAILED DESCRIPTION OF THE INVENTION

The invention concerns truncated $\alpha_2\delta$ -2, $\alpha_2\delta$ -3 or $\alpha_2\delta$ -4 subunit cDNA sequences. These truncated sequences encode soluble secreted polypeptides which retain their affinity for [³H]gabapentin.

Throughout the present specification, the expression "nucleotide sequence" is used to designate indifferently a polynucleotide or a nucleic acid. More precisely, the expression "nucleotide sequence" encompasses the nucleic material and the sequence information and is not restricted to the sequence information (i.e. the succession of letters chosen among the four base letters) that biochemically characterizes a specific DNA or RNA molecule.

As used interchangeably herein, the terms "oligonucleotides", "nucleic acids" and "polynucleotides" include RNA, DNA, or RNA/DNA hybrid sequences of more than one nucleotide in either single chain or duplex form.

Further to its general meaning understood by the one skilled in the art, the term "nucleotide" is also used herein to encompass modified nucleotides which comprise at least one of the following modifications (a) an alternative linking group, (b) an analogous form of purine, (c) an analogous form of pyrimidine, or (d) an analogous sugar. For examples of analogous linking groups, purines, pyrimidines, and sugars, see for example PCT publication N°WO 95/04064.

The polynucleotide sequences of the invention may be prepared by any known method, including synthetic, recombinant, or a combination thereof as well as through any purification methods known in the art.

A) Secreted $\alpha_2\delta$ -2, $\alpha_2\delta$ -3 or $\alpha_2\delta$ -4 subunit polypeptides

The invention comprises polynucleotide sequences encoding a soluble secreted eukaryotic, preferably a soluble secreted mammal $\alpha_2\delta$ -2, $\alpha_2\delta$ -3 or $\alpha_2\delta$ -4 subunit polypeptide. These

sequences particularly include but are not restricted to 1) those sequences encoding a soluble secreted polypeptide of this $\alpha_2\delta$ -2, $\alpha_2\delta$ -3 or $\alpha_2\delta$ -4 subunit which preferably retains its binding affinity for [3 H]gabapentin and 2) nucleotide fragments useful as nucleic acid primers or probes for amplification or detection purposes.

- 5 The expression "soluble secreted $\alpha_2\delta$ -2, $\alpha_2\delta$ -3 or $\alpha_2\delta$ -4 subunit" is intended to designate polypeptide sequences which, when produced by a recombinant host cell, are secreted at least partially into the culture medium rather than remaining associated with the host cell membrane.

10 **1) cDNA fragments encoding soluble secreted $\alpha_2\delta$ -2, $\alpha_2\delta$ -3, $\alpha_2\delta$ -4 subunit polypeptides**

One of the important embodiments of the present invention concerns truncated nucleotide sequences of $\alpha_2\delta$ -2, $\alpha_2\delta$ -3 or $\alpha_2\delta$ -4 subunit cDNAs which encode soluble secreted $\alpha_2\delta$ -2, $\alpha_2\delta$ -3 or $\alpha_2\delta$ -4 subunit polypeptides. The inventors have found that it was possible to
 15 generate deletion mutants of $\alpha_2\delta$ -2, $\alpha_2\delta$ -3 or $\alpha_2\delta$ -4 subunit cDNAs which, when expressed, produce a significant amount of soluble secreted proteins, preferably soluble secreted proteins, which retain their [3 H]gabapentin binding affinity. These truncated nucleotide sequences of the invention are of significant value to the skilled person as they now allow fast and reliable access to significant concentrations of selected soluble secreted $\alpha_2\delta$ -2,
 20 $\alpha_2\delta$ -3 or $\alpha_2\delta$ -4 subunit polypeptides. To that end, the inventors have determined the minimal and optimal fragment lengths required to express a polypeptide which: 1) binds [3 H]gabapentin with sufficient affinity and; 2) is obtained in a soluble secreted form.

The discussion provided below provides comments on possible truncations, giving as an example the human $\alpha_2\delta$ -2, $\alpha_2\delta$ -3 or $\alpha_2\delta$ -4 subunit. However, given the very substantial
 25 cross-species homology for $\alpha_2\delta$ -2, $\alpha_2\delta$ -3 or $\alpha_2\delta$ -4 subunit sequences, the comments below can also be applied to other eukaryotic species, and more particularly other mammalian species such as rat, mouse, rabbit or pig. Their $\alpha_2\delta$ -2, $\alpha_2\delta$ -3 or $\alpha_2\delta$ -4 subunit sequences, which for most are available in public databases, share a very substantial homology with the human $\alpha_2\delta$ -2, $\alpha_2\delta$ -3 or $\alpha_2\delta$ -4 subunit sequences.

30

The inventors believe that the soluble secreted $\alpha_2\delta$ -2 subunit polypeptides which are as close as possible to the native sequence and which are therefore more likely to retain their native folding and hence their [3 H]gabapentin binding properties are those corresponding to the native protein in which amino-acid stretch __1027__ to __the C-terminal end__ of
 35 the amino-acid sequence of SEQ ID N°20 has been deleted. The skilled scientist can quite easily determine within this amino-acid stretch the optimal $\alpha_2\delta$ -2 subunit polypeptides.

The inventors also believe that the soluble secreted $\alpha_2\delta$ -3 subunit polypeptides which are as close as possible to the native sequence and which are therefore more likely to retain

their native folding and hence their $[^3\text{H}]$ gabapentin binding properties are those corresponding to the native protein in which amino-acid stretch ___984___ to _C-terminal end___ of the amino-acid sequence of SEQ ID N°22 has been deleted. The skilled scientist can quite easily determine within this amino-acid stretch the optimal $\alpha_2\delta$ -3 subunit polypeptides.

The invention therefore particularly concerns a nucleotide sequence encoding a polypeptide having at least 80% identity with the polypeptide comprising from amino-acid 1 to between amino-acids _1027___ and _1145___, preferably to between amino-acids ___1062___ and _1145 of SEQ ID N°20.

Preferred nucleotide sequences include those of SEQ ID N°1, SEQ ID N° 2 and SEQ ID N°3.

The invention also concerns a nucleotide sequence encoding a polypeptide having at least 80% identity with the polypeptide comprising from amino-acid 1 to between amino-acids ___984___ and 1085___, preferably to between amino-acids 1019___ and _1085 of SEQ ID N°22.

Preferred nucleotide sequences include those of SEQ ID N°7, SEQ ID N° 8 and SEQ ID N°9.

B) Amplification of the soluble secreted $\alpha_2\delta$ -2, $\alpha_2\delta$ -3 or $\alpha_2\delta$ -4 subunit nucleotide sequences

Another object of the invention consists of a method for the amplification of a nucleic acid encoding a soluble secreted $\alpha_2\delta$ -2, $\alpha_2\delta$ -3 or $\alpha_2\delta$ -4 subunit polypeptide, preferably a polypeptide bearing a $[^3\text{H}]$ gabapentin binding site, said method comprising the steps of:

(a) contacting a test sample suspected of containing the target $\alpha_2\delta$ -2, $\alpha_2\delta$ -3 or $\alpha_2\delta$ -4 subunit nucleic acid, a fragment or a variant thereof, or a sequence complementary thereto, with an amplification reaction reagent comprising a pair of amplification primers which can hybridize under stringent conditions, the $\alpha_2\delta$ -2, $\alpha_2\delta$ -3 or $\alpha_2\delta$ -4 subunit nucleic acid region to be amplified, and

(b) optionally, detecting the amplification products.

The expression $[^3\text{H}]$ gabapentin binding site, when used herein is intended to designate a site which can bind either $[^3\text{H}]$ gabapentin or other ligands such as (S+)-3-isobutyl gaba or (R-)-3-isobutyl gaba.

In a first preferred embodiment of the above method, the nucleic acid encodes a secreted soluble $\alpha_2\delta$ -2, $\alpha_2\delta$ -3 or $\alpha_2\delta$ -4 subunit polypeptide of SEQ ID N°4, SEQ ID N°5, SEQ ID N°6, SEQ ID N°10, SEQ ID N°11, SEQ ID N°12, SEQ ID N°16, SEQ ID N°17 and SEQ ID N°18.

In a second preferred embodiment of the above amplification method, the amplification product is detected by hybridization with a labelled probe having a sequence which is complementary to the amplified region.

5

C) Recombinant vectors and hosts cells for the expression of a secreted soluble $\alpha_2\delta$ -2, $\alpha_2\delta$ -3 or $\alpha_2\delta$ -4 subunit polypeptide

1) Recombinant vectors

10 The present invention also encompasses a family of recombinant vectors comprising any one of the nucleic acids described herein. Firstly, the invention deals with a recombinant vector comprising a nucleic acid selected from the group consisting of:

(a) a purified or isolated nucleic acid encoding a a secreted soluble $\alpha_2\delta$ -2, $\alpha_2\delta$ -3 or $\alpha_2\delta$ -4 subunit having at least 80% amino acid identity with the polypeptide of SEQ ID N°20 or 22, or a sequence complementary thereto;

15 (b) a purified or isolated nucleic acid having at least 90% nucleotide identity with a polynucleotide selected from the group consisting of the nucleotide sequences of SEQ ID N°1, SEQ ID N°2, SEQ ID N°3, SEQ ID No 7, SEQ ID N°8, SEQ ID N°9, SEQ ID No 13, SEQ ID N°14, SEQ ID N°15 or a sequence complementary thereto;

20 (c) a purified or isolated polynucleotide comprising at least 10 consecutive nucleotides of a nucleic acid described in (a) or (b) or a sequence complementary thereto.

In a first preferred embodiment a recombinant vector of the invention is used to amplify the inserted polynucleotide of the invention in a suitable host cell, this polynucleotide being amplified every time the recombinant vector replicates.

25 Recombinant expression vectors comprising a nucleic acid encoding secreted soluble $\alpha_2\delta$ -2, $\alpha_2\delta$ -3 or $\alpha_2\delta$ -4 subunit polypeptides that are described in the present specification are also part of the invention. These include, but are not restricted to, nucleic acids encoding from amino-acid 1 to between amino-acids 1027 and 1145, preferably between amino-acids 1062 and 1145 of SEQ ID N°20, as well as nucleic acids encoding from amino-acid 1 to between amino-acids 984 and 1085, preferably between amino-acids 1019 and 1085, of SEQ ID N°22.

30 Another preferred embodiment of the recombinant vectors according to the invention consist of expression vectors comprising a nucleic acid encoding $\alpha_2\delta$ -2, $\alpha_2\delta$ -3 or $\alpha_2\delta$ -4 subunit polypeptides of the invention, and more preferably a nucleic acid encoding a polypeptide selected from the group consisting of the amino acid sequences of SEQ ID N°4, SEQ ID N°5, SEQ ID N°6, SEQ ID N°10, SEQ ID N°11, SEQ ID N°12, SEQ ID N°16, SEQ ID N°17 and SEQ ID n°18.

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Within certain embodiments, expression vectors can be employed to express the secreted soluble $\alpha_2\delta$ -2, $\alpha_2\delta$ -3 or $\alpha_2\delta$ -4 subunit polypeptides which can then be purified and for example, be used as a immunogen in order to raise specific antibodies directed against said secreted soluble $\alpha_2\delta$ -2, $\alpha_2\delta$ -3 or $\alpha_2\delta$ -4 subunit polypeptides.

Preferred eukaryotic vectors of the invention are listed hereafter as illustrative but not limitative examples: pcDNA3, pFLAG, pCMV-Script, pIND, pMC1NEO, pHIL, pGAPZA, pMT/V5-His-TOPO, pMT/V5-His, pAc5.1/V5-HisA, pDS47/V5-His, pcDNA4, pcDNA6, pEF1, pEF4, pEF6, pUB6, pZeoSV2, pRc/CMV2, pcDM8, pCR3.1, pDisplay, pSecTag2, pVP22, pEMZ, pCMV/Zeo, pSinRep5, pCEP, pREP, pHook-1

Preferred bacteriophage recombinant vectors of the invention are P1 bacteriophage vectors such as described by Sternberg N.L. (1992;1994).

A suitable vector for the expression of a soluble secreted $\alpha_2\delta$ -2, $\alpha_2\delta$ -3 or $\alpha_2\delta$ -4 subunit polypeptide is a baculovirus vector that can be propagated in insect cells and in insect cell-lines. Specific suitable host vectors includes, but are not restricted to :pFastBac-1, pIZ/V5-His, pBacMan-1, pBlueBac4.5, pBlueBacHis2, pMelBacA, pVL1392, pVL1393

The recombinant expression vectors from the invention may also be derived from an adenovirus such as those described by Feldman and Steig. (1996) or Ohno et al. (1994).

Another preferred recombinant adenovirus according to this specific embodiment of the present invention is the human adenovirus type two or five (Ad 2 or Ad 5) or an adenovirus of animal origin (French Patent Application n°FR 93 05 954).

a) Regulatory expression sequences

Expression requires that appropriate signals are provided in the vectors, said signals including various regulatory elements such as enhancers/promoters from both viral and mammalian sources that drive expression of the genes of interest in host cells. The regulatory sequences of the expression vectors of the invention are operably linked to the nucleic acid a soluble secreted $\alpha_2\delta$ -2, $\alpha_2\delta$ -3 or $\alpha_2\delta$ -4 subunit polypeptide.

As used herein, the term “operably linked” refers to a linkage of polynucleotide elements in a functional relationship. For instance, a promoter or an enhancer is operably linked to a coding sequence if it affects the transcription of the coding sequence.

More precisely, two DNA molecules (such as a polynucleotide containing a promoter region and a polynucleotide encoding a desired polypeptide or polynucleotide) are said to be “operably linked” if the nature of the linkage between the two polynucleotides does not : (1) result in the introduction of a frame-shift mutation or (2) interfere with the ability of the polynucleotide containing the promoter to direct the transcription of the coding polynucleotide.

Generally, recombinant expression vectors include origins of replication, selectable markers permitting transformation of the host cell, and a promoter derived from a highly

expressed gene to direct transcription of a downstream structural sequence. The heterologous structural sequence is assembled in an appropriate frame with the translation, initiation and termination sequences, and preferably a leader sequence capable of directing sequences of the translated protein into the periplasmic space or the extra-cellular medium.

In a specific embodiment wherein the vector is adapted for transfecting and expressing desired sequences in eukaryotic host cells, preferred vectors comprise an origin of replication from the desired host, a suitable promoter and an enhancer, and also any necessary ribosome binding sites, polyadenylation site, transcriptional termination sequences, and optionally 5'-flanking non-transcribed sequences.

DNA sequences derived from the SV 40 viral genome, for example SV 40 origin early promoter, enhancer, and polyadenylation sites may be used to provide the required non-transcribed genetic elements.

b) Promoter sequences

Suitable promoter regions used in the expression vectors according to the invention are chosen taking into account the host cell in which the heterologous nucleic acids have to be expressed.

A suitable promoter may be heterologous with respect to the nucleic acid for which it controls the expression, or alternatively can be endogenous to the native polynucleotide containing the coding sequence to be expressed.

Additionally, the promoter is generally heterologous with respect to the recombinant vector sequences within which the construct promoter/coding sequence has been inserted.

Preferred eukaryotic promoters are the CMV, polyhidran or OPIE2.

c) Recombinant host cells

Host cells that have been transformed or transfected with one of the nucleic acids described herein, or with one of the recombinant vector, particularly recombinant expression vector, described herein are also part of the present invention.

Are included host cells that are transformed (prokaryotic cells) or are transfected (eukaryotic cells) with a recombinant vector such as one of those described above. Preferred host cells used as recipients for the expression vectors of the invention are the following:

(a) prokaryotic host cells: *Escherichia coli*, strains. (i.e. DH10 Bac strain) *Bacillus subtilis*, *Salmonella typhimurium* and strains from species such as *Pseudomonas*, *Streptomyces* and *Staphylococcus*;

(b) eukaryotic host cells: HeLa cells (ATCC N°CCL2; N°CCL2.1; N°CCL2.2), Cv 1 cells (ATCC N°CCL70), COS cells (ATCC N°CRL 1650; N°CRL 1651), Sf-9 cells (ATCC N°CRL 1711), C127 cells (ATCC N°CRL-1804), 3T3 cells (ATCC N°CRL-6361),

CHO cells (ATCC N°CCL-61), human kidney 293 cells (ATCC N° 45504; N°CRL-1573), BHK (ECACC N°84100 501; N°84111301), sf 9, sf 21 and hi-5 cells.

D) Production of recombinant secreted soluble $\alpha_2\delta$ -2, $\alpha_2\delta$ -3 or $\alpha_2\delta$ -4 subunit polypeptides

The present invention also concerns a method for producing one of the amino acid sequences described herein and especially a polypeptide selected from the group consisting of the amino acid sequences of SEQ ID N°4, SEQ ID N°5, SEQ ID N°6, SEQ ID N°10, SEQ ID n°11, SEQ ID n°12, SEQ ID n°16, SEQ ID n°17 or SEQ ID n°18 wherein said method comprises the steps of:

(a) inserting the nucleic acid encoding the desired amino acid sequence in an appropriate vector;

(b) culturing, in an appropriate culture medium, a host cell previously transformed or transfected with the recombinant vector of step (a);

(c) harvesting the culture medium thus obtained or lyse the host cell, for example by sonication or osmotic shock;

(d) separating or purifying, from said culture medium, or from the pellet of the resultant host cell lysate, the thus produced recombinant polypeptide of interest.

In some instances, it is required to tag the secreted soluble $\alpha_2\delta$ -2, $\alpha_2\delta$ -3 or $\alpha_2\delta$ -4 subunit polypeptide prior to purification. The tag is then in most instances encoded into the nucleotide sequence that is needed to express the polypeptide. Examples of such tags include, but are not limited to sequences encoding C-myc, FLAG, a sequence of histidine residues, heamagglutinin A, V5, Xpress or GST. Most of these tags can be incorporated directly into the sequence, for instance through PCR amplification by incorporating the appropriate coding sequence in one of the PCR amplification primers. However, the tag can also be introduced by other means such as covalent binding of the appropriate nucleic acid sequence encoding the tag moiety with the 3' or 5' end of the nucleic acid sequence encoding the polypeptide sequence. This is the case for GST.

Purification of the recombinant secreted soluble $\alpha_2\delta$ -2, $\alpha_2\delta$ -3, $\alpha_2\delta$ -4 subunit polypeptides according to the present invention is then carried out by passage onto a nickel or copper affinity chromatography column, such as a Ni NTA column or a Q-Sepharose column.

In another embodiment of the above method, the polypeptide thus produced is further characterized, for example by binding onto an immuno-affinity chromatography column on which polyclonal or monoclonal antibodies directed to the secreted soluble $\alpha_2\delta$ -2 subunit polypeptide of interest have been previously immobilised.

5

In another embodiment of the invention, the secreted soluble $\alpha_2\delta$ -2, $\alpha_2\delta$ -3, $\alpha_2\delta$ -4 subunit polypeptide can be only partially purified. For instance, it can be purified along with other contaminating proteins using an appropriate chromatography matrix such as an ion-exchange chromatography column. In such instances, it is not required to tag the desired polypeptide of interest.

10

The most preferred embodiment contemplated by the inventors concerns the use of a purified tagged secreted soluble $\alpha_2\delta$ -2, $\alpha_2\delta$ -3 or $\alpha_2\delta$ -4 subunit polypeptide. A particularly preferred tag is a nucleotide sequence encoding from 2 to 10, and preferably 6 histidine residues.

15

With regard to the secreted soluble $\alpha_2\delta$ -2, $\alpha_2\delta$ -3 or $\alpha_2\delta$ -4 subunit polypeptide used subsequently in the screening assay of the invention, several possibilities are also open to the skilled person.

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In a first and preferred embodiment, the secreted soluble $\alpha_2\delta$ -2, $\alpha_2\delta$ -3 or $\alpha_2\delta$ -4 subunit polypeptide comprises a tag moiety which can be selected among the tags referred to above. Such tagged polypeptides are particularly useful in SPA or flashplate assays. A preferred tag is the nucleotide sequence encoding histidine residues referred to above.

25

In a second embodiment, the secreted soluble $\alpha_2\delta$ -2, $\alpha_2\delta$ -3 or $\alpha_2\delta$ -4 subunit polypeptide can be used without a tag. This is the case for instance in SPA or flashplate assays comprising beads or plates coated with wheat germ lectin. In such an embodiment, the tag is not needed as the carbohydrate moieties of the secreted soluble $\alpha_2\delta$ -2, $\alpha_2\delta$ -3 or $\alpha_2\delta$ -4 subunit polypeptide bind directly to the wheat germ lectin-coated beads or plates.

30

1) Labelled compounds which bind the secreted soluble $\alpha_2\delta$ -2, $\alpha_2\delta$ -3 or $\alpha_2\delta$ -4 subunit polypeptide

In cases where the $\alpha_2\delta$ -2, $\alpha_2\delta$ -3 or $\alpha_2\delta$ -4 binding site is the [^3H]gabapentin binding site, the preferred labelled compound which can be used is of course gabapentin itself.

- 5 However, gabapentin is not the only labelled compound which can be used in this context. Indeed, it has been previously demonstrated that saturation binding analyses on porcine synaptic plasma cerebral cortex membranes performed in the presence of L-leucine indicate a competitive interaction of the amino acid with the [^3H]gabapentin binding site, significantly reducing [^3H]gabapentin binding affinity for the site. The inventors believe
10 that this competitive interaction is true across across all the amino-acids listed in table 1 below.

15 **TABLE 1**

Binding affinities of selected amino acids ($\text{IC}_{50} < 500\text{nM}$) for the [^3H]gabapentin site in porcine cortical membranes

	<u>COMPOUND</u>	<u>IC_{50} (NM) ARITHMETIC MEAN (N=3) \pm S.E.M.</u>
20	Gabapentin	42.1 ± 5.5
	L-Norleucine	23.6 ± 6.7
	L-Allo-Isoleucine	32.8 ± 6.0
	L-Methionine	49.6 ± 10.0
	L-Leucine	61.3 ± 20.9
25	L-Isoleucine	68.8 ± 1.9
	L-Valine	330 ± 18
	L-Phenylalanine	351 ± 89

- It is therefore possible to use commercially available labelled forms of these high affinity
30 ligands in replacement of gabapentin. The utility of [^3H]L-leucine has been demonstrated in a filter binding assay and in a flashplate assay format. The inventors believe that labelled amino acids but also other compounds, with affinities preferably below 500 nM in the binding assay can be used as replacements of gabapentin.

With regard to the label, several embodiments can be used in the context of the invention. Preferred labels are of course radioactive labels, a list of which is provided further in this specification.

5

2) Assay formats and conditions

Several assay formats can be used to carry out the method of the present invention. Preferred assay formats include scintillation assays such as the scintillation proximity assay (SPA) or the flashplate assay. Other assay formats well known to those skilled in the arts such as the filter binding assay and the centrifugation assay are also contemplated in the present invention.

10

SPA and flashplate assays are preferred assay formats for the present invention. Additional details on these assays are provided below.

15

Scintillation assay format

Scintillation assays technology either involves the use of scintillant beads (for the SPA assay) or plates (for the flashplate assay). SPA beads are usually made from either cerium-doped yttrium ion silicate ($\text{Y}_2\text{SiO}_5:\text{Ce}$) or polyvinyltoluene (PVT) containing an organic scintillant such as PPO. Flashplates commonly used are those such as Ni chelate flashplates although other flashplates can also be used.

20

Assays are usually carried out in aqueous buffers using radioisotopes such as ^3H , ^{125}I , ^{14}C , ^{35}S or ^{33}P that emit low-energy radiation, the energy of which is easily dissipated in an aqueous environment. For example, the electrons emitted by ^3H have an average energy of only 6 keV and have a very short path length ($\sim 1 \mu\text{m}$) in water. If a molecule labelled with one of these isotopes is bound to the bead or flashplate surface, either directly or via interaction with another molecule previously coupled to the bead or flashplate, the emitted radiation will activate the scintillant and produce light. The amount of light produced, which is proportional to the amount of labelled molecules bound to the beads, can be measured conveniently with a liquid scintillation (LS) counter. If the labelled molecule is not attached to the bead or a flashplate surface, its radiation energy is absorbed by the surrounding aqueous solvent before it reaches the bead, and no light is produced. Thus, bound ligands give a scintillation signal, but free ligands do not, and the need for a time-consuming separation step, characteristic of conventional radioligand binding assays, is

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eliminated. The manipulations required in the assays are reduced to a few simple pipetting steps leading to better precision and reproducibility.

The conditions under which SPA and flashplate assays are performed in the context of the present invention are provided below.

5

Scintillation assay conditions

a) SPA assay

The SPA assays is first developed to optimize the conditions under which the radioligand binds the secreted soluble $\alpha_2\delta$ -2, $\alpha_2\delta$ -3 or $\alpha_2\delta$ -4 subunit polypeptide. The parameters which can be varied to optimize radioligand binding in a typical SPA assay using Amersham beads include assay temperature, $\alpha_2\delta$ -2, $\alpha_2\delta$ -3 or $\alpha_2\delta$ -4 subunit polypeptide interaction with the radioligand and the SPA beads, radioligand concentration as well as pH variations.

15 The temperature at which the assay can be carried out can vary from 1 to 30°C. Preferred temperatures range from 18 to 23°C, with 21°C being the most preferred temperature. The interaction of the $\alpha_2\delta$ subunit polypeptide with the SPA beads can be optimized by adjusting the concentration of the polypeptide and by introducing a reagent which will favor this interaction. When 50 mg of Amersham SPA beads are used, the $\alpha_2\delta$ -1 subunit polypeptide concentration may vary from 0.1 to 10 pmoles per well, with the optimal concentration being generally around 5 to 6 pmoles per well.

As for the reagent favoring the interaction between the secreted soluble $\alpha_2\delta$ -2, $\alpha_2\delta$ -3 or $\alpha_2\delta$ -4 subunit polypeptide and the radioligand as well as the Amersham SPA beads, the inventors found that imidazole could be efficiently used for that purpose when the $\alpha_2\delta$ -2, $\alpha_2\delta$ -3 or $\alpha_2\delta$ -4 subunit polypeptide is tagged with an amino acid sequence including 6 histidine residues. Furthermore, and more importantly, it was found that imidazole also enhanced binding of the radioligand to the $\alpha_2\delta$ -2, $\alpha_2\delta$ -3 or $\alpha_2\delta$ -4 polypeptide.

30 The concentration of the radioligand is evaluated with respect to the concentration of secreted soluble $\alpha_2\delta$ -2, $\alpha_2\delta$ -3 or $\alpha_2\delta$ -4 subunit polypeptide present in the assay medium. Generally, the concentration of radioligand varies from 1 nM to 100 nM. A preferred [^3H]gabapentin concentration is about 5 to 20 nM, with a most preferred concentration

being about 10 nM. A preferred [^3H]leucine concentration is also about 5 to 20 nM, with a most preferred concentration being about 10 nM. It is to be noted that the concentration of other radioligands having affinities similar to those of [^3H]gabapentin and [^3H]leucine should also be in the range of about 5 to 20 nM.

5

Once the optimal radioligand binding conditions have been determined, a test ligand can be introduced in the assay medium to evaluate the level of displacement of the radioligand. The concentration of test ligand to be introduced in the assay medium usually varies from 0.1 nM to about 100 μM . A preferred test ligand concentration of about 10 μM is usually a starting point in a high throughput screening assay. Then, depending on the number of hits obtained, it may be lowered or increased.

10

It is to be noted that the parameters set forth above, which have been evaluated for a typical SPA assay using Amersham SPA beads can be adjusted by the skilled person, for example if SPA beads of a different type are used.

15

b) Flashplate assay

Similarly to the SPA assays, the flashplate can first be developed in order to optimize the conditions under which the radioligand binds the $\alpha_2\delta$ subunit polypeptide. The parameters which can be varied to optimize radioligand binding in a typical flashplate assay using NEN Ni chelate flashplates also include assay temperature, secreted soluble $\alpha_2\delta$ -2, $\alpha_2\delta$ -3 or $\alpha_2\delta$ -4 subunit polypeptide interaction with both the radioligand and the flashplates, radioligand concentration as well as pH variations.

20

The temperature at which the assay can be carried out can vary from 1 to 30°C. Preferred temperatures range from 18 to 23°C, with 21°C being the most preferred temperature.

25

The interaction of the secreted soluble $\alpha_2\delta$ -2, $\alpha_2\delta$ -3 or $\alpha_2\delta$ -4 subunit polypeptide with the flashplates can be optimized by adjusting the concentration of the polypeptide and by introducing a reagent which will favor this interaction. When a standard NEN Ni chelate flashplate is used, the secreted soluble $\alpha_2\delta$ -2, $\alpha_2\delta$ -3 or $\alpha_2\delta$ -4 subunit polypeptide volume usually varies between 0.5 and 20 μl for a concentration of secreted soluble $\alpha_2\delta$ -2, $\alpha_2\delta$ -3 or $\alpha_2\delta$ -4 subunit polypeptide of 0.6 pmol/ μl . As the published maximum binding capacity of

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NEN p plates is about 6 pmol per well, the inventors consider that an optimal concentration of secreted soluble $\alpha_2\delta$ -2, $\alpha_2\delta$ -3 or $\alpha_2\delta$ -4 subunit polypeptide is probably around 5 pmol per well at 8 ul.

- 5 With regard to the reagent favoring the interaction between the secreted soluble $\alpha_2\delta$ -2, $\alpha_2\delta$ -3 or $\alpha_2\delta$ -4 subunit polypeptide and the radioligand as well as the flashplates, the inventors believe that imidazole could also be efficiently used for that purpose when the secreted soluble $\alpha_2\delta$ -2, $\alpha_2\delta$ -3 or $\alpha_2\delta$ -4 subunit polypeptide is tagged with an amino acid sequence including 6 histidine residues. The inventors also believe that imidazole
10 concentrations can substantially enhanced binding of the radioligand to the secreted soluble $\alpha_2\delta$ -2, $\alpha_2\delta$ -3 or $\alpha_2\delta$ -4 polypeptide. The optimal concentration of imidazole used to enhance radioligand binding varies depending on the concentration of secreted soluble $\alpha_2\delta$ -2, $\alpha_2\delta$ -3 or $\alpha_2\delta$ -4 subunit polypeptide used in the assay. For instance, when the volume of the $\alpha_2\delta$ -1 subunit polypeptide is about 10 ul ul ($\alpha_2\delta$ -2, $\alpha_2\delta$ -3 or $\alpha_2\delta$ -4
15 polypeptide concentration of 0.6 pmol/ul), the optimal imidazole concentration can vary between 1 and 20 mM, with a concentration of about 10 mM being preferred. As mentioned previously, other compounds such as histidine as well as pH variations may be used to enhance radioligand binding.
- 20 The concentration of the radioligand is evaluated with respect to the concentration of $\alpha_2\delta$ -2, $\alpha_2\delta$ -3 or $\alpha_2\delta$ -4 subunit polypeptide present in the assay medium. Generally, the concentration of radioligand varies from 1 nM to 100 nM. A preferred [^3H]gabapentin concentration is about 5 to 20 nM, with a most preferred concentration being about 10 nM. A preferred [^3H]leucine concentration is also about 5 to 20 nM, with a most preferred
25 concentration being about 10 nM. It is to be noted that the concentration of other radioligands having affinities similar to those of [^3H]gabapentin and [^3H]leucine should also be in the range of about 5 to 20 nM.

- 30 Once the optimal radioligand binding conditions have been determined, a test ligand can be introduced in the assay medium to evaluate the level of displacement of the radioligand. The concentration of test ligand to be introduced in the assay medium usually varies from 0.1 nM to about 100 uM. A preferred test ligand concentration of about 10 uM is usually a

starting point in a high throughput screening assay. Then, depending on the number of hits obtained, it may be lowered or increased.

The inventors have tested the displacement of a particular radioligand, [³H]gabapentin, with (S+)-3-isobutyl gaba. The data provided in the examples which follow clearly shows that the assay can be used in high throughput competition studies.

E) Purified recombinant secreted soluble $\alpha_2\delta$ -2, $\alpha_2\delta$ -3 or $\alpha_2\delta$ -4 polypeptides

Another object of the present invention consists of a purified or isolated recombinant polypeptide comprising the amino acid sequence of a secreted soluble $\alpha_2\delta$ -2, $\alpha_2\delta$ -3 or $\alpha_2\delta$ -4 subunit polypeptide.

Preferred isolated recombinant polypeptides of the invention include those having at least 80%, preferably 90%, more preferably 95, and most preferably 98 or 99%, amino-acid identity with polypeptides comprising from amino acid 1 to between amino-acids 1027 and 1145, preferably between amino-acids 1062 and 1145 of SEQ ID N°20, as well as those having at least 80%, preferably 90%, more preferably 95, and most preferably 98 or 99%, amino-acid identity with polypeptides comprising from amino acid 1 to between amino-acids 984 and 1085, preferably between amino-acids 1019 and 1085 of SEQ ID N°22.

In a further preferred embodiment, the polypeptide comprises an amino acid sequence having at least 80%, preferably 90%, more preferably 95%, and most preferably 98% or 99% amino acid identity with the amino acid sequence of SEQ ID N°4, SEQ ID N°5, SEQ ID N°6, SEQ ID N°10, SEQ ID N°11, SEQ ID N°12, SEQ ID N°16, SEQ ID N°17 and SEQ ID N°19.

F) Modified secreted soluble $\alpha_2\delta$ -2, $\alpha_2\delta$ -3 or $\alpha_2\delta$ -4 subunit polypeptides

The invention also relates to secreted soluble $\alpha_2\delta$ -2, $\alpha_2\delta$ -3 or $\alpha_2\delta$ -4 subunit polypeptide comprising amino acid changes ranging from 1, 2, 3, 4, 5, 10, 20, 25, 30, 35, 40 substitutions, additions or deletions of one amino acid as regards to polypeptides of anyone of the amino acid sequences of the present invention. Preferred sequences are those of SEQ ID N°4, SEQ ID N°5, SEQ ID N°6, SEQ ID N°10, SEQ ID N°11, SEQ ID N°12, SEQ ID N°16, SEQ ID N°17 and SEQ ID N°18.

In the case of an amino acid substitution in the amino acid sequence of a polypeptide according to the invention, one or several consecutive or non-consecutive amino-acids are

replaced by "equivalent" amino-acids. The expression "equivalent" amino acid is used herein to designate any amino acid that may be substituted for one of the amino-acids belonging to the native protein structure without decreasing the binding properties of the corresponding peptides to the antibodies raised against the polypeptides of the invention.

5 In other words, the "equivalent" amino-acids are those which allow the generation or the synthesis of a polypeptide with a modified sequence when compared to the amino acid sequence of the secreted soluble $\alpha_2\delta$ -2, $\alpha_2\delta$ -3 or $\alpha_2\delta$ -4 subunit polypeptides of interest, said modified polypeptide being able to bind to the antibodies raised against the secreted soluble $\alpha_2\delta$ -2, $\alpha_2\delta$ -3 or $\alpha_2\delta$ -4 subunit polypeptide of interest and/or to induce antibodies
10 recognizing the parent polypeptide.

Alternatively, amino acid changes encompassed are those which will not abolish the biological activity of the resulting modified polypeptide. These equivalent amino-acids may be determined either by their structural homology with the initial amino-acids to be replaced, by the similarity of their net charge or of their hydrophobicity, and optionally by
15 the results of the cross-immunogenicity between the parent peptides and their modified counterparts.

The peptides containing one or several "equivalent" amino-acids must retain their specificity and affinity properties to the biological targets of the parent protein, as it can be assessed by a ligand binding assay or an ELISA assay.

20 Examples of amino-acids belonging to specific classes include Acidic (Asp, Glu), Basic (Lys, Arg, His), Non-polar (Ala, Val, Leu, Ile, Pro, Met, Phe, Trp) or uncharged Polar (Gly, Ser, Thr, lys, Tyr, Asn, Gln) amino-acids.

Preferably, a substitution of an amino acid in $\alpha_2\delta$ -2, $\alpha_2\delta$ -3 or $\alpha_2\delta$ -4 subunit polypeptide of the invention, or in a peptide fragment thereof, consists in the replacement of an amino
25 acid of a particular class for another amino acid belonging to the same class.

By an equivalent amino acid according to the present invention is also contemplated the replacement of a residue in the L-form by a residue in the D form or the replacement of a Glutamic acid (E) residue by a Pyro-glutamic acid compound. The synthesis of peptides containing at least one residue in the D-form is, for example, described by Koch (1977).

30 A specific embodiment of a modified peptide of interest according to the present invention, includes, but is not limited to, a peptide molecule, which is resistant to proteolysis. This is a peptide in which the -CONH- peptide bond is modified and replaced by a (CH₂NH) reduced bond, a (NHCO) retro inverso bond, a (CH₂-O) methylene-oxy bond, a (CH₂S) thiomethylene bond, a (CH₂CH₂) carba bond, a (CO-CH₂) cetomethylene bond, a (CHOH-CH₂) hydroxyethylene bond), a (N-N) bound, a E-alcene bond or also a -CH=CH-bond.
35

The invention also encompasses secreted soluble $\alpha_2\delta$ -2, $\alpha_2\delta$ -3 or $\alpha_2\delta$ -4 subunit polypeptide in which at least one peptide bond has been modified as described above.

The polypeptides according to the invention may also be prepared by the conventional methods of chemical synthesis, either in a homogenous solution or in solid phase. As an illustrative embodiment of such chemical polypeptide synthesis techniques, it may be cited the homogenous solution technique described by Houbenweyl (1974).

5 The secreted soluble $\alpha_2\delta$ -2, $\alpha_2\delta$ -3 or $\alpha_2\delta$ -4 subunit polypeptide of interest, or a fragment thereof may thus be prepared by chemical synthesis in liquid or solid phase by successive couplings of the different amino acid residues to be incorporated (from the N-terminal end to the C-terminal end in liquid phase, or from the C-terminal end to the N-terminal end in solid phase) wherein the N-terminal ends and the reactive side chains are previously
10 blocked by conventional groups.

For solid phase synthesis, the technique described by Merrifield (1965a; 1965b) may be used in particular.

G) Antibody production

15 The secreted soluble $\alpha_2\delta$ -2, $\alpha_2\delta$ -3 or $\alpha_2\delta$ -4 subunit polypeptides of the invention and their peptide fragments of interest can be used for the preparation of antibodies.

Polyclonal antibodies may be prepared by immunization of a mammal, especially a mouse or a rabbit, with a polypeptide according to the invention that is combined with an adjuvant
20 of immunity, and then by purifying the specific antibodies contained in the serum of the immunized animal on an affinity chromatography column on which has previously been immobilized the polypeptide that has been used as the antigen.

Monoclonal antibodies may be prepared from hybridomas according to the technique described by Kohler and Milstein (1975).

25 The present invention also deals with antibodies produced by the trioma technique and by the human B-cell hybridoma technique, such as described by Kozbor et al. (1983).

Antibodies of the invention also include chimeric single chain Fv antibody fragments (US Patent N° 4,946,778; Martineau et al., (1998), antibody fragments obtained through phage display libraries Ridder et al. (1995) and humanized antibodies (Leger et al., (1997)).

H) Screening assays

30 The invention concerns a method for the screening of ligands which bind soluble secreted $\alpha_2\delta$ -2, $\alpha_2\delta$ -3 or $\alpha_2\delta$ -4 subunit polypeptide. More particularly, the targeted $\alpha_2\delta$ -2, $\alpha_2\delta$ -3 or $\alpha_2\delta$ -4 subunit binding site is preferably the [^3H]gabapentin binding site. The various
35 parameters of the method of the invention are described in further detail below.

EXAMPLES

5 Example 1

Construction of a nucleotide sequence encoding a soluble secreted human $\alpha_2\delta$ -2 subunit polypeptide deletion mutant of SEQ ID N°23

a) Primer design

10 PCR primers were designed to generate the secreted soluble human $\alpha_2\delta$ -2 deletion mutant of SEQ ID N° 23 as follows:

5' PCR primer: This was designed to engineer in a KOZAK translation initiation consensus sequence prior to the coding sequence (Kozak *JBC* **266** 19867-19870)

15 3' PCR primer: This was designed to engineer in six histidine residues followed by a stop-codon at the desired location in the coding sequence. In addition to the stop codon the $\alpha_2\delta$ -2 primers also included an *Eco* RI restriction site.

The bold region in each primer sequence denotes the 'tagged' region; addition of sequences not present in the template. Primers were custom synthesized by Perkin Elmer

20 Applied Biosystems UK to the ABI ready pure grade, supplied lyophilized then resuspended to 15 μ M in 10mM TE. JB197 and 198 were provided with 5' phosphate groups:

25 5' Primer JB197 (5' -**TCGCCACCATGGCGGTGCCGGCTC**-3' ,SEQ ID N° 25)

3' Primer JB198 (5' -**TCGGAATTCCTCAGTGATGGTGATGGTGATGGGCCCCGCGCCACAGTC**-3' ,SEQ ID N° 28)

b) Protocol for PCR mediated 5' Kozak and 3' 6His tagging of human $\alpha_2\delta$ -2

30

The full length human $\alpha_2\delta$ -2 gene (Gen Bank Accession Number AF042792) in a pcDNA 3 vector as described in Brown, J.P. and Gee,N.S.,Cloning and deletion mutagenesis of the $\alpha_2\delta$ calcium channel subunit from porcine cerebral cortex,*The journal of biological chemistry*, **273**(39):25458-25465) was used as the template in the following PCR reaction.

35 *The reagents were added in the following order in triplicate to a 96 well PCR plate:*

	μ l
10x Pfx Amplification buffer	5
10mM dNTPs	1.5

	50mM MgSO ₄	1	
	15μM JB197	1.5	
	15μM JB198	1.5	
	100ng/μl pcDNA3.1-humans-α ₂ δ-2	1	
5	10x PCR Enhancer	5	
	H ₂ O	32.7	
	2.5 UNITS/μL PFX POLYMERASE		0.8μL

The plate was the cycled on an MJ Tetrad DNA engine according to the following cycling
 10 conditions:

94°C / 2mins
followed by:
 for 30 cycles 94°C / 45sec
 15 58°C / 45sec
 68°C / 4mins
followed by:
 68°C / 10mins
followed by:
 20 hold at 4°C

The 3366bp product was then gel purified from a 1% TAE agarose gel using QIAEX beads and eluted in approximately 50μl TE.

25

Example 2

Cloning of the PCR fragments of Example 1 into the Baculovirus transfer vector pFastBac1

30 The PCR products of Example 1 were cloned into *Stu* I digested, calf intestinal phosphatase dephosphorylated, phenol chloroform extracted and QIAEX gel purified pFastBac1 (Life Technologies) using the Rapid DNA ligation kit (Roche Diagnostics) transforming XL1-blue (α₂δ-1b) *E. Coli* cells:

a) Screening for positive recombinants

Given that the PCR product was cloned by blunt-end ligation a screen was required to select a recombinant with the gene ligated in the positive orientation with respect to the polyhedrin promoter in pFastBac1. This was achieved by restriction digest of miniprep DNA (Qiagen miniprep kit) prepared from colony minicultures and analysis on a 1% TAE agarose gel. A positive clone was identified according to the following digest patterns:

SEQ ID N° 23 in pFastBac1

10 *Eco* RI digest performed on miniprep DNA

	Predicted fragments (bp)
PCR product cloned in a positive orientation	4773 and 3368
PCR product cloned in a negative orientation	8127 and 14

b) Sequencing analysis of selected clones

One positive was selected for this clone and used to prepare a plasmid DNA stock of the desired construct (QIAGEN maxi kit). Confirmatory sequence reactions were performed using the Big Dye terminator sequencing kit and run on an ABI 310 Prism Genetic Analyzer. Sequence analysis of both coding strands was performed using a selection of sequencing oligonucleotide primers.

Example 3

Protocol for establishing baculovirus banks for the expression of the $\alpha_2\delta$ -2 deletion mutant SEQ ID N°23

Essentially, the protocol used to generate the baculovirus banks is that outlined in the Life Technologies Bac-to BacTM baculovirus expression systems manual.

a) Transposition of DH10Bac *E Coli* cells

One ng (5 μ l) of the recombinant pFastBac-1 construct containing the nucleotide sequence encoding the porcine $\alpha_2\delta$ -2 deletion mutant of SEQ ID N°23 was added to 100 μ l of DH10Bac cells thawed on ice. The cells were then mixed gently by tapping the tube then incubated on ice for 30minutes before heat shock treatment by incubation in a 42°C water bath for 45 seconds. The mixture was then chilled on ice for 2 minutes before the addition of 900 μ l of S.O.C. medium. The mixture was then placed in a shaking incubator (200rpm) at 37°C for 4hours. The cells were then serially diluted (10 fold dilutions from 10⁻¹ to 10⁻³)

and 10µl of each dilution plated on LB agar plates containing 50µg/ml kanamycin, 7µg/ml gentamicin, 10µg/ml tetracycline, 100µg/ml Bluo-gal and 40µg/ml IPTG. The plates were incubated at 37°C for between 1 and 3 days until discrete colonies of blue and white colour were discernible.

5

b) Isolation of recombinant DNA

White colonies (containing the recombinant bacmid) were picked and grown for 24 hours (to stationary phase) at 37°C with shaking (200rpm) in 2ml of LB containing 50µg/ml kanamycin, 7µg/ml gentamicin and 10µg/ml tetracycline. 1.5ml of culture was then

10 transferred to a microfuge tube and centrifuged at 14,000xg for 1minute. The supernatant was removed and the cells resuspended gently in 0.3ml of 15mM Tris-HCl (pH8.0), 10mM EDTA, 100µg/ml RNase A. 0.3ml of 0.2N NaOH, 1% SDS was then added and the mixture mixed gently before incubation at 22°C for 5 minutes. Then 0.3ml of 3M Potassium acetate (pH5.5) was added and the sample placed on ice for 10 minutes. After
15 centrifugation at 14,000xg for 10 minutes the supernatant was transferred to a tube containing 0.8ml of isopropanol, mixed then placed on ice for 10 minutes before centrifugation at 14,000xg for 10 minutes. The supernatant was then discarded and the pellet rinsed with 0.5ml of 70% ethanol before centrifugation at 14,000xg for 5 minutes. This 70% ethanol rinse was then repeated before removing all of the supernatant and air
20 drying the pellet for 10minutes at room temperature. The pellet was finally resuspended in 40µl of TE.

c) Transfection of sf9 cells with the recombinant bacmid DNA

A 6-well tissue culture plate was seeded with 0.9×10^6 sf9 cells (cells at log phase having
25 grown from a culture passaged at 0.3×10^6 cells/ml) per 35mm well in 2ml of Sf-900 II SFM media containing 50units/ml penicillin and 50µg/ml streptomycin. Cells were left to attach at 27°C for 1 hour. Bacmid DNA prepared as described above (5µl) was added to 200µl of Sf-900 II SFM media containing 6µl of CELLFECTIN and mixed before incubation at room temperature for 45 minutes. The cells were washed once with 2ml of
30 Sf-900 II SFM media without antibiotics then 0.8ml of Sf-900 II SFM media was added to each tube containing the lipid-DNA complex. The wash buffer was removed from the cells and the 1ml of diluted lipid-DNA complex overlaid on the cells. The cells were incubated for 5hours at 27°C after which time the transfection mixture was removed and 2ml of Sf-

900 II SFM media containing 50units/ml penicillin and 50µg/ml streptomycin was added. The cells were then incubated for 72 hours.

After incubation for 72 hours the media was removed from the cells and centrifuged at 500xg for 5 minutes. The supernatant was then transferred to a fresh tube, this was labelled as the P0 bank and stored at 4°C in the dark. The P1 bank was prepared by passaging sf9 cells at approx 5×10^6 cells/ml to 2×10^6 cells/ml (100ml in a 250ml Erlenmeyer flask) and adding 0.5ml of the P0 bank harvested above. The cells were then incubated shaking (200rpm) at 27°C for 4 days. Under sterile conditions the culture was centrifuged at 500xg for 10 minutes and the supernatant 0.2µM filtered (P1 bank). The P2 bank was prepared by adding 2ml of P1 bank per 400ml culture (in 1L Erlenmeyer flasks) passaged as above to 2×10^6 cells/ml. The culture was incubated as before for 4 days and the supernatant harvested and filtered as described for the P1 bank. The supernatant was first pooled then aliquoted (10ml) and stored at 4°C.

Example 4

Expression of the $\alpha_2\delta$ -2 deletion mutant of SEQ ID N°23

To sf9 cells passaged from $\sim 5 \times 10^6$ cells/ml to 2×10^6 cells/ml in Sf-900 II SFM media was added 0.1ml virus per 100ml of cells of the appropriate viral bank (400ml volumes in 1L Erlenmeyer flasks). The cells were then cultured for 4-5 days at 27°C with 110rpm shaking. Expression of the protein was confirmed by SDS-PAGE and Western blotting using an anti penta-His monoclonal antibody (Qiagen) and was detected in the culture supernatant and cell lysate.

Example 5

Purification of $\alpha_2\delta$ -2deletion mutant of SEQ ID N°23

The- $\alpha_2\delta$ -2 deletion mutant of SEQ ID N°23 was purified from the cell lysate following the purification strategy outlined below:

The culture was centrifuged at 6,000xg for 10 minutes and the supernatant removed. The weight of the cell pellet was determined before re-suspension in 20mM Tris pH8.0, 100mMKCl, 1% P40-Nonidet (100ml per 20g of wet cells). A protease inhibitor cocktail

(Sigma, Cat# P8849), 1ml/L, was added to the mixture. The solution was then stirred for 10 minutes before centrifugation for 1hour at 30,000xg and 4°C. The supernatant was concentrated (30kDa cut off) to approx. ~300ml then centrifuged for 1hour at 100,000xg.

5 Supernatant containing the soluble proteins was diluted 1:3 in 10mM Tris-HCl pH8.0 (equilibration buffer) and loaded onto a pre-equilibrated Q-Sepharose column (2.5cm i.d. x 30cm h.) at a flow rate of 900ml/h. After washing with equilibration buffer until a stable A_{280nm} baseline had been achieved, protein was eluted with 20mM Tris-HCl pH8.0, 0.5M KCl, 10mM Imidazole.

10 The eluate was then loaded onto a Ni-NTA (Qiagen) column (2.5cm i.d. x 6cm h.) pre-equilibrated in 20mM Tris pH8.0, 0.5M KCl, 10mM Imidazole at a flow rate of 2 ml/min. The column was washed successively with buffer A (20mM Tris pH8.0, 0.5M KCl, 20mM Imidazole), buffer B (100mM Tris-HCl pH8.0, 1M KCl), and buffer A again. Elution was
15 performed with buffer C (20mM Tris-HCl pH8.0, 100mM KCl, 0.5M Imidazole). The Ni-NTA eluate (~50ml) was concentrated (30kDa cut-off) to ~2ml and applied at 1ml/min and in 0.2ml aliquots, to an FPLC Superdex-200 column equilibrated in 10mM HEPES, pH7.4, 150mM NaCl. Fractions containing the polypeptide of SEQ ID N°23 were pulled.

20

Example 6

SPA assay of [3 H]gabapentin binding to the secreted soluble human $\alpha_2\delta$ -2 subunit of SEQ ID N°23

25 The assay is carried out at 21°C. Assay components are added in the following order (all reagents are diluted in 10mM HEPES (pH 7.4 at 21°C) to 96-well Optiplates:

	25µl	imidazole at various concentrations (diluted from a 1M stock pH8.0, see assay details)
	50µl	10mM HEPES pH 7.4
30	25µl	(50mg) SPA beads (Amersham)
	100µl	s- $\alpha_2\delta$ -2 subunit polypeptide of SEQ ID No 23 (2µl protein diluted to 100µl)
	25µl	radioligand ([3 H]gabapentin obtained from example 5

Immediately after adding radioligand, the optiplates were loaded in the Packard Top Count scintillation counter to follow the binding time course. Imidazole was first used in the assay to optimize the specific interaction of the protein's 6His tag with the SPA bead. Imidazole itself (up to 100mM) in the filtration assay has no effect on [³H]gabapentin binding (n=1).

Example 7

Ni Flashplate assay of [³H]gabapentin binding to secreted soluble human $\alpha_2\delta$ -2 (SEQ ID N°23)

Assays are carried out at 21°C in a final volume of 250µl in 96-well NEN Ni chelate flash plates. Assay components are added in the following order (all reagents were diluted in 10mM HEPES (pH 7.4 at 21°C)):

- 25µl 10mM HEPES pH7.4
- 25µl imidazole at various concentrations (diluted from a 1M stock pH8.0, see assay details)
- 75µl 10mM HEPES pH 7.4
- 100µl s- $\alpha_2\delta$ -2-6His (2µl protein diluted to 100µl) obtained from example 5
- 25µl radioligand ([³H]gabapentin (65Ci/mmol))

Immediately after adding the radioligand, flash plates are loaded in the Packard Top Count scintillation counter to follow the binding time course. The '[³H] flash plate' programme (cpm) is used to monitor activity. Imidazole is first used in the assay to optimize the specific interaction of the protein's 6His tag with the Ni flashplate.

Example 8

Ni Flashplate assay of [³H]Leucine binding to secreted soluble human $\alpha_2\delta$ -2-6His

The procedure described in example 7 is repeated, except that [³H]gabapentin is replaced by 25 ul (10.1 nM) of [³H]Leucine (141 Ci/mmol).

Example 9**Ni Flashplate assay studying competitive binding of [³H]gabapentin and (S+)-3-isobutyl GABA to human $\alpha_2\delta$ -2-6His (SEQ ID N°23).**

- 5 Assays are carried out at 21°C in a final volume of 250µl in 96-well NEN Ni chelate flash plates. Wells are set up for both 'total' and 'non-specific' binding. Specific binding is defined as that remaining after subtraction of the average of the 'non-specific binding' values from the average of the 'total' binding values. Assay components are added in the following order (all reagents were diluted in 10mM HEPES (pH 7.4 at 21°C)):
- | | | |
|----|----------------------|--|
| 10 | 25µl | 10mM HEPES pH7.4 or 25 ul othe test compound at the appropriate concentration in HEPES |
| | 25µl | 200 mM imidazole (diluted from a 1M stock pH8.0, see assay details) |
| | Total binding | 75µl 10mM HEPES pH 7.4 |
| 15 | Non-specific binding | 50µl 10mM HEPES pH 7.4 and 25µl 100µM (S+)-3-isobutyl GABA |
| | | 100µl $\alpha_2\delta$ -2-6His (2µl protein* diluted to 100µl) |
| | 25µl | radioligand ([³ H]gabapentin or [³ H]Leucine) |
- 20 * The source of $\alpha_2\delta$ -2-6His is that purified by fplc Superdex-200 gel filtration (see example 5)
- Immediately after adding radioligand, flash plates are loaded in the Packard Top Count scintillation counter to follow the binding time course. Incubation time before the assay is 3 hours. The '[³H] flash plate' programme (cpm) is used to monitor activity
- 25 Competition studies are compared across the flash-plate and filter binding methodologies in order to validate the new assay technology with the established filter binding methodology.
- GraphPad Prism software is used to process competition curve data and determine IC₅₀ and
- 30 hill slope values. Twelve point competition curves with half log dilution steps of test compounds are used in the experiments.

Example 10**Filter binding assay of [³H]gabapentin binding to the recombinant polypeptide of SEQ ID N°23**

- 5 Assays were carried out at 21°C in a final volume of 250µl in 96-deep well plates. Assay components were (all reagents were diluted in 10mM HEPES (pH 7.4 at 21°C)):

25µl compound to test

200µl Polypeptide of SEQ ID N°23 (3µl protein diluted to 200µl)

25µl radioligand ([³H]gabapentin (65Ci/mmol))

10

Plates were incubated at room temperature for 1h prior to filtering on to 96-well GF/B Unifilter plates pre-soaked in 0.3% polyethylenimine. Filters were washed with 3x1ml 50mM Tris-HCl (pH 7.4 at 4°C), and dried over-night. Scintillant (Microscint O, 50µl) was added and the plates counted using a Packard Top Count scintillation counter. Specific
 15 binding was ~98% of the 'total' value. In [³H]gabapentin saturation studies, the K_D (nM) obtained was about 10.62. Table 2 below provides the results of the competition studies.

Example 11

- 20 **Construction of a nucleotide sequence encoding a soluble secreted mouse α₂δ-3 deletion mutant of SEQ ID N°25 as follows.**

a) Primer design

- 25 PCR primers were designed to generate the secreted soluble mouse α₂δ-3 deletion mutant of SEQ ID N° 25 as follows:

5' PCR primer: This was designed to engineer in a KOZAK translation initiation consensus sequence prior to the coding sequence (Kozak *JBC* **266** 19867-19870)

3' PCR primer: This was designed to engineer in six histidine residues followed by a stop-codon at the desired location in the coding sequence. In addition to the stop codon the α₂δ-

- 30 3 primers also included an *Eco* RI restriction site.

The bold region in each primer sequence denotes the 'tagged' region; addition of sequences not present in the template. Primers were custom synthesized by Perkin Elmer Applied Biosystems UK to the ABI ready pure grade, supplied lyophilized then

resuspended to 15 μ M in 10mM TE. JB201 and 202 were provided with 5' phosphate groups:

5' Primer JB201 (5'-TCGCCACCATGGCCGGGCCGGGC-3', SEQ ID N°27)

3' Primer JB202 (5'-TCTCAGTGATGGTGATGGTGATGCGATGCACCCCCACACTCTC-3', SEQ ID N°28)

b) Protocol for PCR mediated 5' Kozak and 3' 6His tagging of mouse $\alpha_2\delta$ -3

- 10 The full length mouse $\alpha_2\delta$ -3 gene (Gen Bank Accession number AJ010949) in the pcDNA3 vector as described in Brown, J.P. and Gee, N.S., Cloning and deletion mutagenesis of the $\alpha_2\delta$ calcium channel subunit from porcine cerebral cortex, *The journal of biological chemistry*, **273**(39):25458-25465) was used as the template in the following PCR reaction.

15 *The reagents were added in the following order in triplicate to a 96 well PCR plate:*

	μ l
10x Pfx Amplification buffer	5
10mM dNTPs	1.5
20 50mM MgSO ₄	1
15 μ M JB201	1.5
15 μ M JB202	1.5
100ng/ μ l pcDNA3-mouse- $\alpha_2\delta$ -3	1
10x PCR Enhancer	5
25 H ₂ O	32.7
<u>2.5 UNITS/μL PFX POLYMERASE</u>	<u>0.8μL</u>

The plate was the cycled on an MJ Tetrad DNA engine according to the following cycling conditions:

30 94°C / 2mins

followed by:

for 30 cycles 94°C / 45sec
60°C / 45sec

68°C / 4mins

followed by:

68°C / 10mins

followed by:

5 hold at 4°C

The 3244bp product was then gel purified from a 1% TAE agarose gel using QIAEX beads and eluted in approximately 50µl.

The truncated protein of SEQ ID N°24 was expressed such the procedure of example 2,3
10 and 4.

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SEQUENCE LISTING**1 - human nucleotide sequence**

ATGGCGGTGCCGGCTCGGACCTGCGGCGCCTCTCGGCCCCGGCCCAGCGCGGACTGCGCGCC
 5 CCTGGCCCCGGCTGCGGCCCCCACCCTGGCCCCGGCACCCGGCGCCCCGACGTCCGGGCCCCC
 GCGCCCGCTGTGGCTGCTGC&TGCCGCTTCTACCGCTGCTCGCCGCCCCCGGCGCCTCTGC
 CTACAGCTTCCCCCAGCAGCACACGATGCAGCACTGGGCCCCGGCGTCTGGAGCAGGAGGTC
 GACGGCGTGATGCGGATTTTTGGAGGCGTCCAGCAGCTCCGTGAGATTTACAAGGACAACC
 GGAACCTGTTGAGGTACAGGAGAATGAGCCTCAGAAGTTGGTGAGAAGGTGGCAGGGGA
 10 CATTGAGAGCCTTCTGGACAGGAAGGTGCAGGCCCTGAAGAGACTGGCTGATGCTGCAGAG
 AACTTCCAGAAAGCACACCGCTGGCAGGACAACATCAAGGAGGAAGACATCGTGTACTATG
 ACGCCAAGGCTGACGCTGAGCTGGACGACCCTGAGAGTGAGGATGTGGAAAGGGGTCTAA
 GGCCAGCACCCCTAAGGCTGGACTTCATCGAGGACCCAACTTCAAGAACAAGGTCAACTAT
 TCATACGCGGCTGTACAGATCCCTACGGACATCTACAAAGGCTCCACTGTCATCCTCAATG
 15 AGCTCAACTGGACAGAGGCCCTGGAGAATGTGTTTATGGAAAACCGCAGACAAGACCCAC
 ACTGCTGTGGCAGGTCTTCGGCAGCGCCACAGGAGTCACTCGCTACTACCCGGCCACCCCG
 TGGCGAGCCCCCAAGAAGATCGACCTGTACGATGTCCGAAGGAGACCCTGGTATATCCAGG
 GGGCCTCGTCACCCAAAGACATGGTCATCATCGTGGATGTGAGTGGCAGTGTGAGCGGCCT
 GACCCTGAAGCTGATGAAGACATCTGTCTGCGAGATGCTGGACACGCTGTCTGATGATGAC
 20 TATGTGAATGTGGCCTCGTTCAACGAGAAGGCACAGCCTGTGTTCATGCTTCACACACCTGG
 TGCAGGCCAATGTGCGCAACAAGAAGGTGTTCAAGGAAGCTGTGCAGGGCATGGTGGCCAA
 GGGCACACAGGCTACAAGGCCGGCTTTGAGTATGCCTTTGACCAGCTGCAGAACTCCAAC
 ATCACTCGGGCCAACTGCAACAAGATGATCATGATGTTTACGGATGGTGGTGAGGACCGCG
 TGCAGGACGTCTTTGAGAAGTACAATTGGCCAAACCGGACGGTGCAGCGTGTCTTACTTTCTC
 25 CGTGGGGCAGCATAACTATGACGTCACACCGCTGCAGTGGATGGCCTGTGCCAACAAAGGC
 TACTATTTTGAGATCCCTTCCATCGGAGCCATCCGCATCAACACACAGGAATATCTAGATG
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 30 TGGCTCTGAATGACATCAAGAGGCTGACCCCCAACTACACGCTTGGAGCCAACGGCTATGT
 GTTTGCCATTGACCTGAACGGCTACGTGTTGCTGCACCCCAATCTCAAGCCCCAGACCACC
 AACTTCCGGGAGCCTGTGACTCTGGACTTCCTGGATGCGGAGCTAGAGGATGAGAACAAGG
 AAGAGATCCGTCGGAGCATGATTGATGGCAACAAGGGCCACAAGCAGATCAGAACGTTGGT
 CAAGTCCCTGGATGAGAGGTACATAGATGAGGTGACACGGAACCTACACCTGGGTGCCTATA
 35 AGGAGCACTAACTACAGCCTGGGGCTGGTGCTCCACCCCTACAGCACCTTCTACCTCCAAG
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 10 GATGCTGGCACTCTACAATAACTCCTTCTACACCCGCAAGGAGTCCTATGACTATCAGGCA
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 TGCGAGGCTGGCCGG

2 - human nucleotide sequence

20 ATGGCGGTGCCGGCTCGGACCTGCGGCGCCTCTCGGCCCGGCCAGCGCGGACTGCGCGCC
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 GCGCCCGCTGTGGCTGCTGCTGCCGCTTCTACCGCTGCTCGCCGCCCCCGCGCCTCTGCC
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 25 GAACCTGTTTCGAGGTACAGGAGAATGAGCCTCAGAAGTTGGTGGAGAAGGTGGCAGGGGAC
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 30 CATACGCGGCTGTACAGATCCCTACGGACATCTACAAAGGCTCCACTGTCATCCTCAATGA
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 10 ACTTCCGGGAGCCTGTGACTCTGGACTTCCCTGGATGCGGAGCTAGAGGATGAGAACAAAGGA
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5 TACAGCTTCCCCCAGCAGCACACGATGCAGCACTGGGCCCCGGCGTCTGGAGCAGGAGGTCG
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15 GGCGAGCCCCCAAGAAGATCGACCTGTACGATGTCCGAAGGAGACCCTGGTATATCCAGGG
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30 ACTTCCGGGAGCCTGTGACTCTGGACTTCTGGATGCGGAGCTAGAGGATGAGAACAAAGGA
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 5 GGTGGGCGTCAAGCTGGACCTAGAGGCTTGGGCTGAGAAGTTCAAGGTGCTAGCCAGCAAC
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 15 CAGAGACTGACCAACACCAATCTTCTCTTTGTGGTGGCCGAGAAGCCGCTGTGCAGCCAGT
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 ACAGAAGATACCTCAGACTGTGGCCGCGGGGCC

4 - human amino acid sequence

MAVPARTCGASRPGPARTARPWPGCGPHGPGTRRPTSGPPRPLWLLLPLLPLLAAPGASA
 YSFPQQHTMQHWARRLEQEVDGVMRIFGGVQQLREIYKDNRNLFVQENEPQKLVEKVAGD
 IESLLDRKVQALKRLADAAENFQKAHRWQDNIKEEDIVYYDAKADAELDDPESEDVERGSK
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 25 LLWQVFGSATGVTRYYPATPWRAPKKIDLYDVRRRPWYIQGASSPKDMVIIVDVSGSVSGL
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 YEDALGLGLVVTGTLPLVFNLTDGPGGEKKNQLILGVMGIDVALNDIKRLTPNYTLGANGYV
 30 FAIDLNGYVLLHPNLKPQTTNFREPVTLDFLDAELEDENKEEIRRS MIDGNKGHKQIRTLV
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 ESEGHVFIAPREYCKDLNASDNNTFLKNFIELMEKVTPDSKQCNFLLHNLILDTGITQQ
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 35 RTHQDQPQKCGPNSHCEMDCEVNNE DLLCVLIDDGGFLVLSNQNHQWDQVGRFFSEVDANL
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 QLLYGLIYHSWFQADPAEAE GSPETRESSCVMKQTQYYFGSVNASYNAIIDCGNCSRLFHA
 QRLTNTNLLFVVAEKPLCSQCEAGR

5 - human amino acid sequence

MAVPARTCGASRPGPARTARPWPGCGPHPGPGTRRPTSGPPRPLWLLLPLLPLLAAPGASA
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 IESLLDRKVQALKRLADAAENFQKAHRWQDNIKEEDIVYYDAKADAELDDPESEDVERGSK
 5 ASTLRRLDFIEDPNFKNKVNYSYAAVQIPTDIYKGSTVILNELNWTEALENVFMENRRQDPT
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 TLKLMKTSVCEMLDLSDDDYVNVASFNEKAQPVSCFTHLVQANVRNKKVFKEAVQGMVAK
 GTTGYKAGFEYAFDQLQNSNITRANCNKMIMMFTDGGEDRVQDVFEKYNWPNRTVRVFTFS
 VGQHNYDVTPLQWMACANKGYFFEIPSIGAIRINTQEYLDVLGRPMVLAGEAKQVQWNTV
 10 YEDALGLGLVVTGTLPVFNLTDGPGGEKKNLILGVMGIDVALNDIKRLTPNYTLGANGYV
 FAIDLNGYVLLHPNLKPQTTNFREPVTLDFLDAELEDENKEEIRRSMIDGNKGHKQIRTLV
 KSLDERYIDEVTRNYTWVPIRSTNYSLGLVLPPYSTFYQLANLSDQILQVKYFEFLLPSSF
 ESEGHVFIAPREYCKDLNASDNNTFLKNFIELMEKVTPDSKQCNNFLLHNLILDTGITQQ
 LVERVWRDQDLNTYSLLAVFAATDGGITRVFPNKAEDWTENPEPFNASFYRRSLDNHGYV
 15 FKPPHQDALLRPLELENDTVGILVSTAVELSLGRRTLRAVVGVKLDLEAWAEKFKVLASN
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 MLALYNNSFYTRKESYDYQAACAPQPPGNLGAAPRGVFPVPTVADFLNLAWWTSAAAWSLFQ
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6 - human amino acid sequence

MAVPARTCGASRPGPARTARPWPGCGPHPGPGTRRPTSGPPRPLWLLLPLLPLLAAPGASA
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 25 ASTLRRLDFIEDPNFKNKVNYSYAAVQIPTDIYKGSTVILNELNWTEALENVFMENRRQDPT
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 30 YEDALGLGLVVTGTLPVFNLTDGPGGEKKNLILGVMGIDVALNDIKRLTPNYTLGANGYV
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TEDTSDCGRGA

7 - human nucleotide sequence

5 ATGGCCGGGCGGGCTCGCCGCGCCGCGCTCCCGGGGGCCTCGGCGCTTCTCGCTGCCG
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 15 AAGGAGACTACAGGGAATATTGCTTGTGAAGACTGCTCCAAGTCCTTTGTTCATCCAGCAAA
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 CCCCATC

8 - human nucleotide sequence

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 GCAGCATGAAAGGACTCCGTCTGACTATCGCGAAGCAAACAGTCTCATCCATTTTGGATAC
 ACTTGGGGATGATGACTTCTTCAACATAATTGCTTATAATGAGGAGCTTCACTATGTGGAA
 35 CCTTGCCTGAATGGAACCTTTGGTGCAGCCGACAGGACAAACAAAGAGCACTTCAGGGAGC
 ATCTGGACAAACCTTTTCGCCAAAGGAATTGGAATGTTGGATATAGCTCTGAATGAGGCCTT
 CAACATTCTGAGTGATTTCAACCACACGGGACAAGGAAGTATCTGCAGTCAGGCCATCATG
 CTCATAACTGATGGGGCGGTGGACACCTATGATACAATCTTTGCAAAATACAATTGGCCAG

ATCGAAAGGTTTCGCATCTTCACATACCTCATTGGACGAGAGGCTGCGTTTGCAGACAATCT
 AAAGTGGATGGCCTGTGCCAACAAAGGATTTTTTACCCAGATCTCCACCTTGGCTGATGTG
 CAGGAGAATGTCATGGAATACCTTCACGTGCTTAGCCGGCCCAAAGTCATCGACCAGGAGC
 ATGATGTGGTGTGGACCGAAGCTTACATTGACAGCACTCTGACTGATGATCAGGGCCCCGT
 5 CCTGATGACCACTGTAGCCATGCCTGTGTTTAGTAAGCAGAACGAAACCAGATCGAAGGGC
 ATTCTTCTGGGAGTGGTTGGCACAGATGTCCCAGTGAAAGAACTTCTGAAGACCATCCCCA
 AATACAAGTTAGGGATTACGGTTATGCCTTTGCAATCACAAATAATGGRTATATCCTGAC
 GCATCCGGAACCTCAGGCTGCTGTACGAAGAAGGAAAAAGCGAAGGAAACCTAACTATAGT
 AGCGTTGACCTCTCTGAGGTGGAGTGGGAAGACCGAGATGACGTGTTGAGAAATGCTATGG
 10 TGAATCGAAAGACGGGGAAGTTTTCCATGGAGGTGAAGAAGACAGTGGACAAAGGGAAACG
 GGTTTTGGTGATGACAAATGACTACTATTATACAGACATCAAGGGTACTCCTTTCAGTTTA
 GGTGTGGCGCTTTCCAGAGGTCATGGGAAATATTTCTTCCGAGGGAATGTAACCATCGAAG
 AAGGCCTGCATGACTTAGAACATCCCGATGTGTCCTTGGCAGATGAATGGTCCTACTGCAA
 CACTGACCTACACCCTGAGCACCGCCATCTGTCTCAGTTAGAAGCGATTAAGCTCTACCTA
 15 AAAGGCAAAGAACCTCTGCTCCAGTGTGATAAAGAATTGATCCAAGAAGTCCTTTTTTGACG
 CGGTGGTGAGTGCCCCCATTTGAAGCGTATTGGACCAGCCTGGCCCTCAACAAATCTGAAAA
 TTCTGACAAGGGCGTGGAGGTTGCCTTCCTCGGCACTCGCACGGGCCTCTCCAGAATCAAC
 CTGTTTGTGCGGGCTGAGCAGCTCACCAATCAGGACTTCCTGAAAGCTGGCGACAAGGAGA
 ACATTTTTTAACGCAGACCATTTCCTCTCTGGTACCGAAGAGCCGCTGAGCAGATTCCAGG
 20 GAGCTTCGTCTACTCGATCCCATTCAGCACTGGACCAGTCAATAAAAGCAATGTGGTGACA
 GCAAGTACATCCATCCAGCTCCTGGATGAACGGAAATCTCCTGTGGTGGCAGCTGTAGGCA
 TTCAGATGAAACTTGAATTTTTCCAAAGGAAGTTCTGGACTGCCAGCAGACAGTGTGCTTC
 CCTGGATGGCAAATGCTCCATCAGCTGTGATGATGAGACTGTGAATTGTTACCTCATAGAC
 AATAATGGATTTATTTTGGTGTCTGAAGACTACACACAGACTGGAGACTTTTTTGGTGAGA
 25 TCGAGGGAGCTGTGATGAACAAATTGCTAACAATGGGCTCCTTTAAAAGAATTACCCTTTA
 TGACTACCAAGCCATGTGTAGAGCCAACAAGGAAAGCAGCGATGGCGCCCATGGCCTCCTG
 GATCCTTATAATGCCCTTCCTCTCTGCAGTAAAATGGATCATGACAGAACTTGTCTTGTTCC
 TGGTGGAATTTAACCTCTGCAGTTGGTGGCACTCCGATATGACAGCTAAAGCCCAGAAATT
 GAAACAGACCCTGGAGCCTTGTGATACTGAATATCCAGCATTCTGTCTCTGAGCGCACCATC
 30 AAGGAGACTACAGGGAATATTGCTTGTGAAGACTGCTCCAAGTCCTTTGTCATCCAGCAAA
 TCCCAAGCAGCAACCTGTTTCATGGTGGTGGTGGACAGCAGCTGCCTCTGTGAATCTGTGGC
 CCCATCACCATGGCACCCATTGAAATCAGGTATAATGAATCCCTTAAGTGTGAACGTCTA
 AAG

35 **9 - human nucleotide sequence**

ATGGCCGGGCGGGCTCGCCGCGCCGCGCTCCCGGGGGGCTCGGCGCTTCTCGCTGCCG
 CGCTTCTCTACGCCGCGCTGGGGGACGTGGTGCGCTCGGAGCAGCAGATACCGCTCTCCGT
 GGTGAAGCTCTGGGCCTCGGCTTTTGGTGGGGAGATAAAATCCATTGCTGCTAAGTACTCC

GGTTCCCAGCTTCTGCAAAAGAAATACAAAGAGTATGAGAAAGACGTTGCCATAGAAGAAA
TTGATGGCCTCCAAGTGGTAAAGAAGCTGGCAAAGAACATGGAAGAGATGTTTTCACAAGAA
GTCTGAGGCCGTCAGGCGTCTGGTGGAGGCTGCAGAAGAAGCACACCTGAAACATGAATTT
GATGCAGACTTACAGTATGAATACTTCAATGCTGTGCTGATAAATGAAAGGGACAAAGACG
5 GGAATTTTTTTGGAGCTGGGAAAGGAATTCATCTTAGCCCCAAATGACCATTTTAATAATTT
GCCTGTGAACATCAGTCTAAGTGACGTCCAAGTACCAACGAACATGTACAACAAAGACCCCT
GCAATTGTCAATGGGGTTTATTGGTCTGAATCTCTAAACAAAGTTTTTTGTAGATAACTTTG
ACCGTGACCCATCTCTCATATGGCAGTACTTTGGAAGTGCAAAGGGCTTTTTTTAGGCAGTA
TCCGGGGATTAAATGGGAACCAGATGAGAATGGAGTCATTGCCTTCGACTGCAGGAACCGA
10 AAATGGTACATCCAGGCAGCAACTTCTCCGAAAGACGTGGTCATTTTAGTTGACGTCAGTG
GCAGCATGAAAGGACTCCGTCTGACTATCGCGAAGCAAACAGTCTCATCCATTTTGGATAC
ACTTGGGGATGATGACTTCTTCAACATAATTGCTTATAATGAGGAGCTTCACTATGTGGAA
CCTTGCCTGAATGGAACTTTGGTGCAAGCCGACAGGACAAACAAAGAGCACTTCAGGGAGC
ATCTGGACAAACTTTTCGCCAAAGGAATTGGAATGTTGGATATAGCTCTGAATGAGGCCTT
15 CAACATTCTGAGTGATTTCAACCACACGGGACAAGGAAGTATCTGCAGTCAGGCCATCATG
CTCATAACTGATGGGGCGGTGGACACCTATGATACAATCTTTGCAAATACAATTGGCCAG
ATCGAAAGGTTGCGATCTTACATACCTCATTGGACGAGAGGCTGCGTTTGCAGACAATCT
AAAGTGGATGGCCTGTGCCAACAAAGGATTTTTTTACCCAGATCTCCACCTTGGCTGATGTG
CAGGAGAATGTCATGGAATACCTTCACGTGCTTAGCCGGCCCAAAGTCATCGACCAGGAGC
20 ATGATGTGGTGTGGACCGAAGCTTACATTGACAGCACTCTGACTGATGATCAGGGCCCCGT
CCTGATGACCACTGTAGCCATGCCTGTGTTTAGTAAGCAGAACGAAACCAGATCGAAGGGC
ATTCTTCTGGGAGTGGTTGGCACAGATGTCCAGTGAAAGAACTTCTGAAGACCATCCCCA
AATACAAGTTAGGGATTACGGTTATGCCTTTGCAATCACAAATAATGGRTATATCCTGAC
GCATCCGGAACCTCAGGCTGCTGTACGAAGAAGGAAAAAAGCGAAGGAAACCTAACTATAGT
25 AGCGTTGACCTCTCTGAGGTGGAGTGGGAAGACCGAGATGACGTGTTGAGAAATGCTATGG
TGAATCGAAAGACGGGGAAGTTTTCCATGGAGGTGAAGAAGACAGTGGACAAAGGGAAACG
GGTTTTGGTGATGACAAATGACTACTATTATACAGACATCAAGGGTACTCCTTTTCACTTTA
GGTGTGGCGCTTTCCAGAGGTCATGGGAAATATTTCTTCCGAGGGAATGTAACCATCGAAG
AAGGCCTGCATGACTTAGAACATCCCGATGTGTCTTGGCAGATGAATGGTCCTACTGCAA
30 CACTGACCTACACCCTGAGCACCGCCATCTGTCTCAGTTAGAAGCGATTAAAGCTCTACCTA
AAAGGCAAAGAACCTCTGCTCCAGTGTGATAAAGAATTGATCCAAGAAGTCCTTTTTTGACG
CGGTGGTGAGTGCCCCCATTTGAAGCGTATTGGACCAGCCTGGCCCTCAACAAATCTGAAAA
TTCTGACAAGGGCGTGGAGGTTGCCTTCTCGGCACTCGCACGGGCCTCTCCAGAATCAAC
CTGTTTGTGCGGGGCTGAGCAGCTACCAATCAGGACTTCTTGAAAGCTGGCGACAAGGAGA
35 ACATTTTTTAACGCAGACCATTTCCCTCTCTGGTACCGAAGAGCCGCTGAGCAGATTCCAGG
GAGCTTCGTCTACTCGATCCCATTCAGCACTGGACCAGTCAATAAAAGCAATGTGGTGACA
GCAAGTACATCCATCCAGCTCCTGGATGAACGGAAATCTCCTGTGGTGGCAGCTGTAGGCA
TTCAGATGAAACTTGAATTTTTTCAAAGGAAGTTCTGGACTGCCAGCAGACAGTGTGCTTC

CCTGGATGGCAAATGCTCCATCAGCTGTGATGATGAGACTGTGAATTGTTACCTCATAGAC
 AATAATGGATTTATTTTGGTGTCTGAAGACTACACACAGACTGGAGACTTTTTTGGTGAGA
 TCGAGGGAGCTGTGATGAACAAATTGCTAACAATGGGCTCCTTTAAAAGAATTACCCTTTA
 TGACTACCAAGCCATGTGTAGAGCCAACAAGGAAAGCAGCGATGGCGCCCATGGCCTCCTG
 5 GATCCTTATAATGCCTTCCTCTCTGCAGTAAAATGGATCATGACAGAACTTGTCTTGTTC
 TGGTGGGAATTTAACCTCTGCAGTTGGTGGCACTCCGATATGACAGCTAAAGCCCAGAAATT
 GAAACAGACCCTGGAGCCTTGTGATACTGAATATCCAGCATTCTGTCTCTGAGCGCACCATC
 AAGGAGACTACAGGGAATATTGCTTGTGAAGACTGCTCCAAGTCCTTTGTCATCCAGCAAA
 TCCCAAGCAGCAACCTGTTTCATGGTGGTGGTGGACAGCAGCTGCCTCTGTGAATCTGTGGC
 10 CCCCATCACCATGGCACCCATTGAAATCAGGTATAATGAATCCCTTAAGTGTGAACGTCTA
 AAGGCCCAGAAGATCAGAAGGGCCCAGAAGATCAGAAGGCGCCCAGAATCTTGTTCATGGCT
 TCCATCCTGAGGAGAATGCAAGGGAGTGTGGGGGTGCGCCG

10 - human amino acid sequence

15 MAGPGSPRRASRGASALLAAALLYAALGDVVRSEQQIPLSVVKLWASAFGGEIKSIAAKYS
 GSQLLQKKYKEYEKDVAIEEIDGLQLVKKLAKNMEEMFHKKSEAVRRLVEAAEEAHLKHEF
 DADLQYEFYFNAVLINERDKDGNFLELGKEFILAPNDHFNNLPVNISLSDVQVPTNMYNKDP
 AIVNGVYWSESLNKVFVDNFDNRDPSLIWQYFGSAKGFFRQYPGIKWEPDENGVIADFCDNR
 KWIQAATSPKDVVILVDVSGSMKGLRLTIAKQTVSSILDTLGDDDDFNIIAYNEELHYVE
 20 PCLNGTLVQADRTNKEHFREHLDKLFAKGIGMLDIALNEAFNILSDFNHTGQGSICSQAIM
 LITDGAVDTYDTIFAKYNWPDRKVRIFTYILIGREAAAFADNLKWMACANKGFFTQISTLADV
 QENVMEYLHVLSRPKVIDQEHDDVVWTEAYIDSTLTDDQGPVLMTTVAMPVFSKQNETRSKG
 ILLGVVGTDPVKELLKTIPKYKLGIGHYAFAITNNGYILTHPELRLLYEEGKKRRKPNYS
 SVDLSEVEWEDRDDVLRNAMVNRKTGKFSMEVKKTVDKGRVLVMTNDYYYTDIKGTPFSL
 25 GVALSRGHGKYFFRGNVTIEEGLHDLHPDVSLADEWSYCNLDLHPEHRHLSQLEAIKLYL
 KGKEPLLQCDKELIQEVLFDVVSAPIEAYWTSALNKSSENSDKGVEVAFLGTRTGLSRIN
 LFGVGAQLTNQDFLKAGDKENIFNADHFPLWYRRAAEQIPGSFVYSIPFSTGPNKSNVVT
 ASTSIQLLDERKSPVVAAVGIQMKLEFFQRKFWTASRQCASLDGKCSISCDDTVNICYLID
 NNGFILVSEDYTQTGDFFGIEGAVMNKLLTMGSFKRITLYDYQAMCRANKESSDGAHGLL
 30 DPYNAFLSAVKWIMTELVLFLVEFNLCSWWHSDMTAKAQKLKQTLPCDTEYPAFVSERTI
 KETTGNIACEDCSKSFVIQQIPSSNLFMVVVDSSCLCESVAPI

11 - human amino acid sequence

MAGPGSPRRASRGASALLAAALLYAALGDVVRSEQQIPLSVVKLWASAFGGEIKSIAAKYS
 35 GSQLLQKKYKEYEKDVAIEEIDGLQLVKKLAKNMEEMFHKKSEAVRRLVEAAEEAHLKHEF
 DADLQYEFYFNAVLINERDKDGNFLELGKEFILAPNDHFNNLPVNISLSDVQVPTNMYNKDP
 AIVNGVYWSESLNKVFVDNFDNRDPSLIWQYFGSAKGFFRQYPGIKWEPDENGVIADFCDNR
 KWIQAATSPKDVVILVDVSGSMKGLRLTIAKQTVSSILDTLGDDDDFNIIAYNEELHYVE

PCLNGTLVQADRTNKEHFREHLDKLFAKGIGMLDIALNEAFNILSDFNHTGQGSICSQAIM
 LITDGAVDITYDTIFAKYNWPDRKVRIFTYILIGREAAAFADNLKWMACANKGFFTQISTLADV
 QENVMEYLHVLSRPKVIDQEHDVVWTEAYIDSTLTDDQGPVLMTTVAMPVFSKQNETRSKG
 ILLGVVGTDVPVKELLKTIPKYKLGIGHGYAFAITNNGYILTHPELRLLYEKGKKRRKPNYS
 5 SVDLSEVEWEDRDDVLRNAMVNRKTGKFSMEVKKTVDKGKRVLMVTNDYYYTDIKGTPFSL
 GVALSRGHGKYFFRGNVTIEEGLHDLEHPDVSLADEWSYCNTDLHPEHRHLSQLEAIKLYL
 KGKEPLLQCDKELIQEVLFDVVSAPIEAYWTSALNKSSENSDKGVEVAFLGTRTGLSRIN
 LFVGAEQLTNQDFLKAGDKENIFNADHFPLWYRRAAEQIPGSFVYSIPFSTGPNKSNVVT
 ASTSIQLLDERKSPVVAAVGIQMKLEFFQRKFWTASRQCASLDGKCSISCDDETVCNYLID
 10 NNGFILVSEDYDTQTGDFGFEIEGAVMNKLLTMGSFKRITLYDYQAMCRANKESSDGAHGLL
 DPYNAFLSAVKWIMTELVLFLVEFNLCSSWWHSDMTAKAQKLKQTLPCDTEYPAFVSERTI
 KETTGNIACEDCSKSFVIQQIPSSNLFMVVVDSSCLCESVAPITMAPIEIRYNESLKCERL
 K

12 - human amino acid sequence

MAGPGSPRRASRGASALLAALLYAALGDVVRSEQQIPLSVVKLWASAFGGEIKSIAAKYS
 GSQLLQKKYKEYEKDVAIEEIDGLQLVKKLAKNMEEMFHKKSEAVRRLVEAAEEAHLKHEF
 DADLQYEFYNAVLINERDKDGNFLELGKEFILAPNDHFNNLPVNISLSVDVQVPTNMYNKDP
 AIVNGVYWSESLNKVFVDNFDNRDPSLIWQYFGSAKGFFRQYPGIKWEPDENGVIADFCDNR
 20 KWIYQAATSPKDVVILVDVSGSMKGLRLTIKQTVSSILDTLGDDDDFNIIAYNEELHYVE
 PCLNGTLVQADRTNKEHFREHLDKLFAKGIGMLDIALNEAFNILSDFNHTGQGSICSQAIM
 LITDGAVDITYDTIFAKYNWPDRKVRIFTYILIGREAAAFADNLKWMACANKGFFTQISTLADV
 QENVMEYLHVLSRPKVIDQEHDVVWTEAYIDSTLTDDQGPVLMTTVAMPVFSKQNETRSKG
 ILLGVVGTDVPVKELLKTIPKYKLGIGHGYAFAITNNGYILTHPELRLLYEKGKKRRKPNYS
 25 SVDLSEVEWEDRDDVLRNAMVNRKTGKFSMEVKKTVDKGKRVLMVTNDYYYTDIKGTPFSL
 GVALSRGHGKYFFRGNVTIEEGLHDLEHPDVSLADEWSYCNTDLHPEHRHLSQLEAIKLYL
 KGKEPLLQCDKELIQEVLFDVVSAPIEAYWTSALNKSSENSDKGVEVAFLGTRTGLSRIN
 LFVGAEQLTNQDFLKAGDKENIFNADHFPLWYRRAAEQIPGSFVYSIPFSTGPNKSNVVT
 ASTSIQLLDERKSPVVAAVGIQMKLEFFQRKFWTASRQCASLDGKCSISCDDETVCNYLID
 30 NNGFILVSEDYDTQTGDFGFEIEGAVMNKLLTMGSFKRITLYDYQAMCRANKESSDGAHGLL
 DPYNAFLSAVKWIMTELVLFLVEFNLCSSWWHSDMTAKAQKLKQTLPCDTEYPAFVSERTI
 KETTGNIACEDCSKSFVIQQIPSSNLFMVVVDSSCLCESVAPITMAPIEIRYNESLKCERL
 KAQKIRRRPESCHGFHPEENARECGGAP

13 - human nucleotide sequence

AGTGGCCTCCTGAGAAGCAGCTTGTTTCGTGGGCTCCGAGAAGGTCTCCGACAGGAAGTTCC
 TGACACCTGAGGACGAGGCCAGCGTGTTACCCCTGGACCGCTTCCCGCTGTGGTACCGCCA
 GGCCTCAGAGCATCCTGCTGGCAGCTTCGTCTTCAACCTCCGCTGGGCAGAAGGACCAGAA

AGTGC GGGTGAACCCATGGTGGTGACGGCAAGCACAGCTGTGGCGGTGACCGTGGACAAGA
 GGACAGCCATTGCTGCAGCCGCGGGCGTCCAAATGAAGCTGGAATTCCTCCAGCGCAAATT
 CTGGGCGGCAACGCGGCAGTGCAGCACTGTGGATGGGCCGTGCACACAGAGCTGCGAGGAC
 AGTGATCTGGACTGCTTCGTCAATCGACAACAACGGGTTTCATTCTGATCTCCAAGAGGTCCC
 5 GAGAGACGGGAAGATTTCTGGGGGAGGTGGATGGTGCTGTCCTGACCCAGCTGCTCAGCAT
 GGGGGTGTTTCAGCCAAGTGACTATGTATGACTATCAGGCCATGTGCAAACCCTCGAGTCAC
 CACCACAGTGCAGCCCAGCCCCCTGGTCAGCCCAATTTCTGCCTTCTTGACGGCGACCAGGT
 GGCTGCTGCAGGAGCTGGTGCTGTTCTGCTGGAGTGGAGTGTCTGGGGCTCCTGGTACGA
 CAGAGGGGCGGAGGCCAAAAGTGTTCTTCCATCACTCCCAACAACACAAGAAGCAGGACCCG
 10 CTGCAGCCCTGCGACACGGAGTACCCCGTGTTCTGTGTACCAGCCGGCCATCCGGGAGGCCA
 ACGGGATCGTGGAGTGCGGGCCCTGCCAGAAGGTATTTGTGGTGCAGCAGATTCCCAACAG
 TAACCTCCTCCTCCTGGTGACAGACCCACCTGTGACTGCAGCATCTTCCCACCAAGTG

14 - human nucleotide sequence

15 AGTGGCCTCCTGAGAAGCAGCTTGTTCTGTGGGCTCCGAGAAGGTCTCCGACAGGAAGTTCC
 TGACACCTGAGGACGAGGCCAGCGTGTTCAACCTGGACCGCTTCCCGCTGTGGTACCGCCA
 GGCCTCAGAGCATCCTGCTGGCAGCTTCGTCTTCAACCTCCGCTGGGCAGAAGGACCAGAA
 AGTGC GGGTGAACCCATGGTGGTGACGGCAAGCACAGCTGTGGCGGTGACCGTGGACAAGA
 GGACAGCCATTGCTGCAGCCGCGGGCGTCCAAATGAAGCTGGAATTCCTCCAGCGCAAATT
 20 CTGGGCGGCAACGCGGCAGTGCAGCACTGTGGATGGGCCGTGCACACAGAGCTGCGAGGAC
 AGTGATCTGGACTGCTTCGTCAATCGACAACAACGGGTTTCATTCTGATCTCCAAGAGGTCCC
 GAGAGACGGGAAGATTTCTGGGGGAGGTGGATGGTGCTGTCCTGACCCAGCTGCTCAGCAT
 GGGGGTGTTTCAGCCAAGTGACTATGTATGACTATCAGGCCATGTGCAAACCCTCGAGTCAC
 CACCACAGTGCAGCCCAGCCCCCTGGTCAGCCCAATTTCTGCCTTCTTGACGGCGACCAGGT
 25 GGCTGCTGCAGGAGCTGGTGCTGTTCTGCTGGAGTGGAGTGTCTGGGGCTCCTGGTACGA
 CAGAGGGGCGGAGGCCAAAAGTGTTCTTCCATCACTCCCAACAACACAAGAAGCAGGACCCG
 CTGCAGCCCTGCGACACGGAGTACCCCGTGTTCTGTGTACCAGCCGGCCATCCGGGAGGCCA
 ACGGGATCGTGGAGTGCGGGCCCTGCCAGAAGGTATTTGTGGTGCAGCAGATTCCCAACAG
 TAACCTCCTCCTCCTGGTGACAGACCCACCTGTGACTGCAGCATCTTCCCACCAAGTGCTG
 30 CAGGAGGCGACAGAAGTCAAATATAATGCCTCTGTCAAATGTGACCGGATGCGC

15 - human nucleotide sequence

AGTGGCCTCCTGAGAAGCAGCTTGTTCTGTGGGCTCCGAGAAGGTCTCCGACAGGAAGTTCC
 TGACACCTGAGGACGAGGCCAGCGTGTTCAACCTGGACCGCTTCCCGCTGTGGTACCGCCA
 35 GGCCTCAGAGCATCCTGCTGGCAGCTTCGTCTTCAACCTCCGCTGGGCAGAAGGACCAGAA
 AGTGC GGGTGAACCCATGGTGGTGACGGCAAGCACAGCTGTGGCGGTGACCGTGGACAAGA
 GGACAGCCATTGCTGCAGCCGCGGGCGTCCAAATGAAGCTGGAATTCCTCCAGCGCAAATT
 CTGGGCGGCAACGCGGCAGTGCAGCACTGTGGATGGGCCGTGCACACAGAGCTGCGAGGAC

AGTGATCTGGACTGCTTCGTCATCGACAACAACGGGTTCAATTCTGATCTCCAAGAGGTCCC
 GAGAGACGGGAAGATTTCTGGGGGAGGTGGATGGTGCTGTCCTGACCCAGCTGCTCAGCAT
 GGGGGTGTTTCAGCCAAGTGACTATGTATGACTATCAGGCCATGTGCAAACCCTCGAGTCAC
 CACCACAGTGCAGCCCAGCCCCTGGTCAGCCCAATTTCTGCCTTCTTGACGGCGACCAGGT
 5 GGCTGCTGCAGGAGCTGGTGCTGTTCTCTGCTGGAGTGGAGTGTCTGGGGCTCCTGGTACGA
 CAGAGGGGCGGAGGCCAAAAGTGTCTTCCATCACTCCCAACAACAAGAAGCAGGACCCG
 CTGCAGCCCTGCGACACGGAGTACCCCGTGTTCTGTGTACCAGCCGGCCATCCGGGAGGCCA
 ACGGGATCGTGGAGTGCGGGCCCTGCCAGAAGGTATTTGTGGTGCAGCAGATTCCCAACAG
 TAACCTCCTCCTCCTGGTGACAGACCCACCTGTGACTGCAGCATCTTCCCACCAGTGCTG
 10 CAGGAGGCGACAGAAGTCAAATATAATGCCTCTGTCAAATGTGACCGGATGCGCTCCCAGA
 AGCTCCGCCGGCGACCAGACTCCTGCCACGCCTTCCATCCAGAGGAGAATGCCCAGGACTG
 CGGCGGCGCCTCG

16 - human amino acid sequence

15 SGLLRSSLFVGSEKVS DRKFLTPED EASVFTLDRFPLWYRQASEHPAGSFVFNLRWAEGPE
 SAGEPMVVTASTAVAVTVDKRTAIAAAAGVQMKLEFLQRKFWAATRQCSTVDGPCTQSCED
 SDLD CFVIDNNGFILISKRSRETGRFLGEVDGAVLTQLLSMGVFSQVTMYDYQAMCKPSSH
 HHSAAQPLVSPISAFILTATRWLLQELVLFLEWSVWGSWYDRGAEAKSVFHSHKHKKQDP
 LQPCDTEYPVFVYQPAIREANGIVECGPCQKV FVVQQIPNSNLLLLVTDPTCDCSIFPPV

17 - human amino acid sequence

SGLLRSSLFVGSEKVS DRKFLTPED EASVFTLDRFPLWYRQASEHPAGSFVFNLRWAEGPE
 SAGEPMVVTASTAVAVTVDKRTAIAAAAGVQMKLEFLQRKFWAATRQCSTVDGPCTQSCED
 SDLD CFVIDNNGFILISKRSRETGRFLGEVDGAVLTQLLSMGVFSQVTMYDYQAMCKPSSH
 25 HHSAAQPLVSPISAFILTATRWLLQELVLFLEWSVWGSWYDRGAEAKSVFHSHKHKKQDP
 LQPCDTEYPVFVYQPAIREANGIVECGPCQKV FVVQQIPNSNLLLLVTDPTCDCSIFPPVL
 QEATEVKYNASVKCDRMR

18 - human amino acid sequence

30 SGLLRSSLFVGSEKVS DRKFLTPED EASVFTLDRFPLWYRQASEHPAGSFVFNLRWAEGPE
 SAGEPMVVTASTAVAVTVDKRTAIAAAAGVQMKLEFLQRKFWAATRQCSTVDGPCTQSCED
 SDLD CFVIDNNGFILISKRSRETGRFLGEVDGAVLTQLLSMGVFSQVTMYDYQAMCKPSSH
 HHSAAQPLVSPISAFILTATRWLLQELVLFLEWSVWGSWYDRGAEAKSVFHSHKHKKQDP
 LQPCDTEYPVFVYQPAIREANGIVECGPCQKV FVVQQIPNSNLLLLVTDPTCDCSIFPPVL
 35 QEATEVKYNASVKCDRMRSQKLRRRPDSCHAFHPEENAQDCGGAS

19 - human alpha2 delta-2 nucleotide sequence

CGGGCAGCGCAGCCCGCAGAGGCGCTGCGGGCCCGTGAGCCCCGGAGGCCCCTCGCGGAGA
 AGGCGGCGGCGGAGGAGAGGCCGAGTTACCGCCCGCCGCCCGCGCCCCCAACCCGCGCG
 CCGCCGCGCGCCGCCACTGCCCCCCTCCCCGCGGCGCCGCATCTTGAATGGAAACATG
 5 GCGGTGCCGGCTCGGACCTGCGGCGCCTCTCGGCCCGGCCAGCGCGGACTGCGCGCCCCT
 GGCCCCGCTGCGGCCCCACCTGGCCCCGGCACCCGGCGCCCGACGTCCGGGCCCCCGCG
 CCCGCTGTGGCTGCTGCTGCCGCTTCTACCGCTGCTCGCCGCCCGGCGCCTCTGCCTAC
 AGCTTCCCCCAGCAGCACACGATGCAGCACTGGGCCCCGGCGTCTGGAGCAGGAGGTCGACG
 GCGTGATGCGGATTTTGGAGGCGTCCAGCAGCTCCGTGAGATTTACAAGGACAACCGGAA
 10 CCTGTTTCGAGGTACAGGAGAATGAGCCTCAGAAGTTGGTGGAGAAGGTGGCAGGGGACATT
 GAGAGCCTTCTGGACAGGAAGGTGCAGGCCCTGAAGAGACTGGCTGATGCTGCAGAGAACT
 TCCAGAAAGCACACCGCTGGCAGGACAACATCAAGGAGGAAGACATCGTGTAATGACGC
 CAAGGCTGACGCTGAGCTGGACGACCCTGAGAGTGAGGATGTGGAAAGGGGGTCTAAGGCC
 AGCACCCCTAAGGCTGGACTTCATCGAGGACCCAACTTCAAGAACAAGGTCAACTATTTCAT
 15 ACGCGGCTGTACAGATCCCTACGGACATCTACAAAGGCTCCACTGTCATCCTCAATGAGCT
 CAACTGGACAGAGGCCCTGGAGAATGTGTTTCATGAAAACCGCAGACAAGACCCACACTG
 CTGTGGCAGGTCTTCGGCAGCGCCACAGGAGTCACTCGCTACTACCCGGCCACCCCGTGGC
 GAGCCCCCAAGAAGATCGACCTGTACGATGTCCGAAGGAGACCCTGGTATATCCAGGGGGC
 CTCGTCACCCAAAGACATGGTCATCATCGTGGATGTGAGTGGCAGTGTGAGCGGCCCTGACC
 20 CTGAAGCTGATGAAGACATCTGTCTGCGAGATGCTGGACACGCTGTCTGATGATGACTATG
 TGAATGTGGCCTCGTTCAACGAGAAGGCACAGCCTGTGTTCATGCTTCACACACCTGGTGCA
 GGCCAATGTGCGCAACAAGAAGGTGTTCAAGGAAGCTGTGCAGGGCATGGTGGCCAAGGGC
 ACCACAGGCTACAAGGCCGGCTTTGAGTATGCCTTTGACCAGCTGCAGAACTCCAACATCA
 CTCGGGCCAACTGCAACAAGATGATCATGATGTTACGGATGGTGGTGAGGACCGCGTGCA
 25 GGACGTCTTTGAGAAGTACAATTGGCCAAACCGGACGGTGCGCGTGTTTACTTTCTCCGTG
 GGGCAGCATAACTATGACGTACACCCGCTGCAGTGGATGGCCTGTGCCAACAAGGCTACT
 ATTTTGAGATCCCTTCCATCGGAGCCATCCGCATCAACACACAGGAATATCTAGATGTGTT
 GGGCAGGCCCATGGTGTGCTGGCAGGCAAGGAGGCCAAGCAGGTTTCAGTGGACCAACGTGTAT
 GAGGATGCACTGGGACTGGGGTTGGTGGTAACAGGGACCCTCCCTGTTTTCAACCTGACAC
 30 AGGATGGCCCTGGGAAAAGAAGAACAGCTGATCCTGGGCGTGATGGGCATTGACGTGGC
 TCTGAATGACATCAAGAGGCTGACCCCCAACTACACGCTTGGAGCCAACGGCTATGTGTTT
 GCCATTGACCTGAACGGCTACGTGTTGCTGCACCCCAATCTCAAGCCCCAGACCACCAACT
 TCCGGGAGCCTGTGACTCTGGACTTCCTGGATGCGGAGCTAGAGGATGAGAACAAGGAAGA
 GATCCGTGCGGAGCATGATTGATGGCAACAAGGGCCACAAGCAGATCAGAACGTTGGTCAAG
 35 TCCCTGGATGAGAGGTACATAGATGAGGTGACACGGAATAACCTGGGTGCCTATAAGGA
 GCACTAACTACAGCCTGGGGCTGGTGCTCCACCCCTACAGCACCTTCTACCTCCAAGCCAA
 TCTCAGTGACCAGATCCTGCAGGTCAAGTATTTTGAGTTCCTGCTCCCCAGCAGCTTTGAG
 TCTGAAGGACACGTTTTTCATTGCTCCCAGAGAGTACTGCAAGGACCTGAATGCCTCAGACA

ACAACACCGAGTTTCCTGAAAACTTTATTGAGCTCATGGAGAAAGTGACTCCAGACTCCAA
 GCAGTGCAACAACCTTCCTTCTGCACAACCTGATCTTGGACACGGGCATCACGCAGCAGCTG
 GTAGAGCGTGTGTGGAGGGACCAGGATCTCAACACGTACAGCCTACTGGCCGTGTTTCGCTG
 CCACAGACGGTGGCATCACCCGAGTCTTCCCCAACAAGGCAGCTGAGGACTGGACAGAGAA
 5 CCCTGAGCCCTTCAATGCCAGCTTCTACCGCCGCAGCCTGGATAACCACGGTTATGTCTTC
 AAGCCCCCACACCAGGATGCCCTGTTAAGGCCGCTGGAGCTGGAGAATGACACTGTGGGCA
 TCCTCGTCAGCACAGCTGTGGAGCTCAGCCTAGGCAGGCGCACACTGAGGCCAGCAGTGGT
 GGGCGTCAAGCTGGACCTAGAGGCTTGGGCTGAGAAGTTCAAGGTGCTAGCCAGCAACCGT
 ACCCACCAAGACCAGCCTCAGAAGTGC GGCCCCAACAGCCACTGTGAGATGGACTGCGAGG
 10 TTAACAATGAGGACTTACTCTGTGTCTCATTGATGATGGAGGATTCTTGGTGCTGTCAAA
 CCAGAACCATCAGTGGGACCAGGTGGGCAGGTTCTTCAGTGAGGTGGATGCCAACCTGATG
 CTGGCAGCTCTACAATAACTCCTTCTACACCCGCAAGGAGTCTTATGACTATCAGGCAGCCT
 GTGCCCCCTCAGCCCCCTGGCAACCTGGGTGCTGCACCCCGGGGTGTCTTTGTGCCACCGT
 TGCAGATTTCTTAACCTGGCCTGGTGACCTCTGCTGCCGCCTGGTCCCTGTTCCAGCAG
 15 CTTCTCTACGGCCTCATCTACCACAGCTGGTTCCAAGCAGACCCCGCGGAGGCCGAGGGGA
 GCCCCGAGACGCGCGAGAGCAGCTGCGTCATGAAACAGACCCAGTACTACTTCGGCTCGGT
 AAACGCCTCCTACAACGCCATCATCGACTGCGGAAACTGCTCCAGGCTGTTCCACGCGCAG
 AGACTGACCAACACCAATCTTCTCTTTGTGGTGGCCGAGAAGCCGCTGTGCAGCCAGTGCG
 AGGCTGGCCGGCTGCTGCAGAAGGAGACGCACTGCCAGCGGACGGCCCCGAGCAGTGTGA
 20 GCTAGTGCAGAGACCGCGATACCGGAGAGGCCCGCACATCTGCTTCGACTACAACGCGACA
 GAAGATACTCAGACTGTGGCCGCGGGGCCCTCCTTCCCGCCGTCGCTGGGCGTCCTGGTCT
 CCCTGCAACTGCTGCTCCTCCTGGGCCTGCCGCCCCGGCCGAGCCTCAAGTCTCGTCCA
 CGCCTCTCGCCGCTCTGAGCACCTTGCCCCACCCACCTCCACTCCCACCTCACCCGGCC
 TCTTCGCCTTTCCACCCTCCTGCCCCACACTCCCCGCCTTAGAGCCTCGTCCCTCCCTCA
 25 CTGAAGGACCTGAGCTGGCCAGGCCCTGAGAGTCTGGTCTGCGCCTTGGGATGGGGAGTCC
 CAAAGCGGGACGCCGCAGGTGTTTGGCACCCAAATCACATCTCACCTCCGAACTGTTCAAG
 TGTCCCCAGACCTTCTTGCCCTGCTGGGCTCCCCCAGTGGGATGGGACAGGGAGGCCACA
 CGCACTGGTGCCAAAACCAGGCCTCTGCTGCCGCCCTTCCTGGAGGCTGCCTATGTTGGGG
 GGGACCCTGCCTCAGCTGACCCGGCCTCTCTGCCCCACCCAAGCCCAAACCTTGTTTCTGT
 30 GAGAATAGTGGAGGAAGGTGAGATGGCCAGTTTGAAGCCTGTGCCTCCCAGCTTAAATCCT
 AGCAGGAGAGAGGCTCTGGGGCAGCCCCCATGGGCTCCTGCCCTTTTCAGGCCTACAGCCA
 CATCCCCAAGCCCACCAGGTGTCAGGATAGTCACAGTGATAACAGTTCAGACACTACCCCA
 TATACACCTGGAACATTGAGGATGGAACTGGACTCACATTCGACATACCCCACTGGGCAC
 ACGCACAAACACACACACTATGGGGTGGGGTGGGTGTAGGGGCTTACAAAGCCTTACACAG
 35 GGCGAGGGGTTGGTGGGAGGGTTGGCACCTGCACACTCCATCTCCTGCTCACCACCTGCCT
 CTAATCTGAGCTGCAGCCTGGCTGGTCTCTCCCATTTCTAAAGCTGAATGTCAAACAGTGCC
 AAATGCTGGGGCAGGGGTGAAGAACCCTCTGTCCCACCCCTAGCCACCAGTGTCTCCAA
 GTGCCCCCTCACCTCTCCAGGTGCTCATTGTAACCATTTCTCACTAGTGTGAGCCCCCAG

TGGGACCACATGCCACTGCCTGCACCTTTCGGCAGAGGAACCCCCACCAGACATCACCCCTT
 TGCCTTAGCAGGGGTGACTTTGTCTCTCCTGGCTGGGCCATCCTTCCGCCAATCTGGCCCT
 TACACACTCAGGCCTGTGCCCCACTCCCTATCTCCTTCCCACCCCTACACACACACTCCCTG
 CTTGCAGGAGGCCAAACTGTCCCTCCCTTGCTGAACACACACACACACACACACACAGG
 5 TGGGGACTGGGCACAGCTCTTCACACCATTTCATTCTGGTCATTTCCCCCAAAGGCATCCCA
 GCCTGGGGGGCCAGTGGGGAACTGAGGGCAAGGGGATATAGTGATGGGGCTCAGATGGACTG
 GGAGGAGGGGGAGGGTGATGCATTAATTAATGGCTTCGTTAATTAATGTCATGTTGCTTGT
 CGCTTTCTCAGTGTGTGTGTGTGGTCCATGCCCACTGCTGGTGCCAGGGTGGGTGTCCATG
 TGCACCCGGCCTGGATGCCAGCTGTGTCTTTCGGGGGCGTGCGTGTAAGTGTAGTGTAGTC
 10 AGGTGCTCAATGGAGAATATAAACATATACAGAAAAATATATATTTTAAGTTTAAAAACA
 GAAAAACAGACAAAACAATCCCCATCAGGTAGCTGTCTAACCCCCAGCTGGGTCTAATCCT
 TCTCATTACCCACCCGACCTGGCTGCCCCCTCACCTTGGGCTGGGGGACTGGGGGGCCATTT
 CCTTTTCTCTGCCCTTTTTTTTGTGTTCTATTTTGTACAGACAAGTTGGAAAAACAACAGC
 GACAAAAAAGTCAAGAACTTTGTAAAATATCGTGTGTGTGATTCTTGTAAAATATTTTC
 15 AAATGGTTTATTACAGAAGATCAGTTATTAAATAATGTTTCATATTTTCACTTC

20- human alpha2 delta-2 amino acid sequence

MAVPARTCGASRPGPARTARWPWPGCGPHPGPGTRRPTSGPPRPLWLLLPLLLAAPGASA
 YSFPQQHTMQHWARRLEQVDGVMRIFGGVQQLREIYKDNRNLFVQENEPQKLVEKVAGD
 20 IESLLDRKVQALKRLADAAENFQKAHRWQDNIKEEDIVYYDAKADAELDDPESEDVERGSK
 ASTLRDLDFIEDPNFKNKVNSYAAVQIPTDIYKGSTVILNELNWTEALENVFMENRRQDPT
 LLWQVFGSATGVTRYYPATPWRAPKIDLYDVRPPWYIQGASSPKDMVIIIDVSGSVSGL
 TLKLMKTSVCEMLDTLSDDDYVNVASFNEKAQPVSCFTHLVQANVRNKKVFKEAVQGMVAK
 GTTGKAGFEYAFDQLQNSNITRANCNKMIMMFTDGGEDRVQDVFEKYNWPNRTRVRVFTFS
 25 VGQHNYDVTPLQWMAKNGYYFEIPSIGAIRINTQEYLDVLGRPMVLGKEAKQVQWTVN
 YEDALGLGLVVTGTLPVFNLTQDGPGEKKNQLILGVMGIDVALNDIKRLTPNYTLGANGYV
 FAIDLNGYVLLHPNLKPQTTFNREPVTLDFLDAELEDENKEEIRSMIDGNKGHKQIRTLV
 KSLDERYIDEVTRNYTWVPIRSTNYSGLVLPPYSTFYLVQANLSDQILQVKYFEFLLPSSF
 ESEGHVFIAPREYCKDLNASDNNTEFLKNFIELMEKVTPDSKQCNNFLLHNLILDTGITQQ
 30 LVERVWRDQDLNTYSLLAVFAATDGGITRVFPNKAEDWTENPEPFNASFYRRSLDNHGYV
 FKPPHQDALLRPLELENDTVGILVSTAVELSLGRRTLPAVVGVKLDLEAWAEKFKVLASN
 RTHQDQPQKCGPNSHCEMDCEVNNEALLCVLIDDGGFLVLSNQNHQWDQVGRFFSEVDANL
 MLALYNNSFYTRKESYDYQAACAPQPPGNLGAAPRGVFVPTVADFLNLAWWTSAAAWSLFQ
 QLLYGLIYHSWFQADPAEAEKSPETRESSCVMKQTQYYFGSVNASYNAIIDCGNCSRLFHA
 35 QRLTNTNLLFVVAEKPLCSQCEAGRLLQKETHCPADGPEQCELVQRPRYRRGPHICFDYNA
 TEDTSDCGRGASFPPSLGVLVSLQLLLLLGLPPRPQPQVLVHASRRL

TACTATAGGGCGGCCGCGAATTCGGCACGAGGCGGCGCGGAGCGGAGCAGGCAGCCCCGCG

GGCCCTCAACAAATCTGAAAATTCTGACAAGGGCGTGGAGGTTGCCTTCCTCGGCACTCGC
 ACGGGCCTCTCCAGAATCAACCTGTTTGTGCGGGCTGAGCAGCTCACCAATCAGGACTTCC
 TGAAAGCTGGCGACAAGGAGAACATTTTAAACGCAGACCATTTCCTCTCTGGTACCGAAG
 AGCCGCTGAGCAGATTCCAGGGAGCTTCGTCTACTCGATCCCATTGAGCACTGGACCAGTC
 5 AATAAAAGCAATGTGGTGACAGCAAGTACATCCATCCAGCTCCTGGATGAACGGAAATCTC
 CTGTGGTGGCAGCTGTAGGCATTGAGATGAACTTGAATTTTTCCAAAGGAAGTTCTGGAC
 TGCCAGCAGACAGTGTGCTTCCCTGGATGGCAAATGCTCCATCAGCTGTGATGATGAGACT
 GTGAATTGTTACCTCATAGACAATAATGGATTTATTTTGGTGTCTGAAGACTACACACAGA
 CTGGAGACTTTTTTGGTGAGATCGAGGGAGCTGTGATGAACAAATTGCTAACAATGGGCTC
 10 CTTTAAAAGAATTACCTTTTATGACTACCAAGCCATGTGTAGAGCCAACAAGGAAAGCAGC
 GATGGCGCCCATGGCCTCCTGGATCCTTATAATGCCTTCCTCTCTGCAGTAAATGGATCA
 TGACAGAACTTGTCTTGTTCCTGGTGGAATTTAACCTCTGCAGTTGGTGGCACTCCGATAT
 GACAGCTAAAGCCCAGAAATTGAAACAGACCCTGGAGCCTTGTGATACTGAATATCCAGCA
 TTCGTCTCTGAGCGCACCATCAAGGAGACTACAGGGAATATTGCTTGTGAAGACTGCTCCA
 15 AGTCCTTTGTGATCCAGCAAATCCCAAGCAGCAACCTGTTGATGGTGGTGGTGGACAGCAG
 CTGCCTCTGTGAATCTGTGGCCCCCATCACCATGGCACCCATTGAAATCAGGTATAATGAA
 TCCCTTAAGTGTGAACGTCTAAAGGCCAGAAAGATCAGAAGGCGCCAGAACTTTGTGATG
 GCTTCCATCCTGAGGAGAAATGCAAGGGAGTGTGGGGGTGCGCCGAGTCTCCAAGCCCAGAC
 AGTCCTCCTTCTGCTCCCTCTGCTTTTGTGATGCTCTTCTCAAGGTGACACTGACTGAGATGT
 20 TCTCTTACTGACTGAGATGTTCTCTTGGCATGCTAAATCATGGATAAACTGTGAACCAAAA
 TATGGTGCAACATACGAGACATGAATATAGTCCAACCATCAGCATCTCATCATGATTTTAA
 ACTGTGCGTGATATAAACTCTTAAAGATATGTTGACAAAAAGTTATCTATCATCTTTTTAC
 TTTGCCAGTCATGCAAATGTGAGTTTGCCACATGATAATCACCTTTCATCAGAAATGGGAC
 CGCAAGTGGTAGGCAGTGTCCCTTCTGCTTGAAACCTATTGAAACCAATTTAAACTGTGT
 25 ACTTTTTTAAATAAAGTATATTAAATCATAAAAAAAAAAAAAAAAAAAAAA

22 - human alpha2 delta-3 amino acid sequence

MAGPGSPRRASRGASALLAAALLYAALGDVVRSEQQIPLSVVKLWASAFGGEIKSIAAKYS
 GSQLLQKKYKEYEKDVAIEEIDGLQLVKKLAKNMEEMFHKKSEAVRRLVEAAEEAHLKHEF
 30 DADLQYEFNAVLINERDKDGNFLELGKEFILAPNDHFNNLPVNISLSDVQVPTNMYNKDP
 AIVNGVYWSESLNKVFVDNFDRLPSLIWQYFGSAKGFFRQYPGIKWEPDENGVIADFCDNR
 KWIQAATSPKDVVILVDVSGSMKGLRLTIKQTVSSILDTLGDDDFNIIAYNEELHYVE
 PCLNGTLVQADRTNKEHFREHLDKLFAKGIGMLDIALNEAFNLSDFNHTGQGSICSQAIM
 LITDGAVDITYDTIFAKYNWPDRKVRIFTYLIGREAAAFADNLKWMACANKGFFTQISTLADV
 35 QENVMEYLHVLSRPKVIDQEHVWTEAYIDSTLTDDQGPVLMTTVAMPVFSKQNETRSKG
 ILLGVVGTDPVKELLKTIPKYKLGIGHYAFAITNNGYILTHPELRLLYEEGKKRRKPNYS
 SVDLSEVEWEDRDDVLRNAMVNRKTGKFSMEVKKTVDKGKRVLMVTNDYYYTDIKGTPFSL
 GVALSRGHGKYFFRGNVTIEEGLHDLEHPDVSLADEWSYCNTDLHPEHRHLSQLEAIKLYL

KGKEPLLQCDKELIQEVLFDVVSAPIEAYWTSALNKSSENSDKGVEVAFLGTRTGLSRIN
 LFVGAEQLTNQDFLKAGDKENIFNADHFPLWYRRAAEQIPGSFVYSIPFSTGPVNKSNVVT
 ASTSIQLLDERKSPVVAAVGIQMKLEFFQQRKFWTASRQCASLDGKCSISCDDETVNCYLID
 NNGFILVSEDYTQTGDFGEIEGAVMNKLLTMGSFKRITLYDYQAMCRANKESSDGAHGLL
 5 DPYNAFLSAVKWIMTELVLFLVEFNLCSSWWHSDMTAKAQKLKQTLPCDTEYPAFVSERTI
 KETTGNIACEDCSKSFVIQQIPSSNLFMVVVDSSCLCESVAPITMAPIEIRYNESLK CERL
 KAQKIRRRPESCHGFHPEENARECGGAPSLQAQTVLLLLLPLLLMLFSR

23 - human amino acid sequence

10 MAVPARTCGASRPGPARTARWPWPGCGPHPGPGTRRPTSGPPLWLLLPLLLAAPGASA
 YSFPQQHTMQHWARRLEQEVDGVMRIFGGVQQLREIYKDNRNLFVQENEPQKLVEKVAGD
 IESLLDRKVQALKRLADAAENFQKAHRWQDNIKEEDIVYYDAKADAELDDPESEDVERGSK
 ASTLRDLFIEDPNFKNKVNSYAAVQIPTDIYKGSTVILNELNWTEALENVFMENRRQDPT
 LLWQVFGSATGVTRYYPATPWRAPKKIDLYDVRRRPWYIQGASSPKDMVIIVDVSGSVSGL
 15 TLKLMKTSVCEMLD TLSDDDYVNVASFNEKAQPVSCFTHLVQANVRNKKVFKEAVQGMVAK
 GTTGKAGFEYAFDQLQNSNITRANCNKMIMMFTDGGEDRVQDVFEKYNWPNRTRVFTFS
 VGQHNYDVTPLQWMACANKGYFEIPSIGAIRINTQEYLDVLGRPMVLAGEAKQVQWTVN
 YEDALGLGLVVTGTLPVFNLTDGPGGEKKNLILGVMGIDVALNDIKRLTPNYTLGANGYV
 FAIDLNGYVLLHPNLKPQTTNFREPVTLDFLDAELEDENKEEIRRS MIDGNKGHKQIRTLV
 20 KSLDERYIDEVTRNYTWVPIRSTNYSLGLVLPPYSTFYLVQANLSDQILQVKYFEFLLPSSF
 ESEGHVFIAPREYCKDLNASDNNTFELKNFIELMEKVTPDSKQCNNFLLHNLILDGTITQQ
 LVERVWRDQDLNTYSLLAVFAATDGGITRVFPNKAAEDWTENPEPFNASFYRRSLDNHGYV
 FKPPHQDALLRPLELENDTVGILVSTAVELSLGRRTLRAVVGVKLDLEAWAEKFKVLASN
 RTHQDQPQKCGPNSHCEMDCEVNNE DLCLVLDIDGGFLVLSNQNHQWDQVGRFFSEVDANL
 25 MLALYNNSFYTRKESYDYQAACAPQPPGNLGAAPRGVFPVPTVADFLNLAWWTSAAAWSLFQ
 QLLYGLIYHSWFQADPAEAGSPETRESSCVMKQTQYYFGSVNASYNAIIDCGNCSRLFHA
 QRLTNTNLLFVVAEKPLCSQCEAGRLLQKETHCPADGPEQCELVQRPRYRRGPHICFDYNA
 TEDTSDCGRGAHHHHH

30 24- mouse amino acid sequence

MAGPGSLCCASRGASALLATALLYAALGDVVRSEQQIPLSVVKLWASAFGGEIKSIAAKYS
 GSQLLQKKYKEYEKDVAIEEIDGLQLVKKLAKIMEEMFHKKSEAVRRLVEAAEEAHLKHEF
 DADLQYEFNAVLINERDKDGNFLELGKEFILAPNDHFNNLPVNISLSDVQVPTNMYNKDP
 AIVNGVYWSESLNKVFVDNFD RDP SLIWQYFGSAKGFFRQYPGIKWEPDENGVI AFDCNRN
 35 KWIQAATSPKDVVILVDVSGSMKGLRLTIKQTVSSILDTLGDDDFNIIITYNEELHYVE
 PCLNGTLVQADRTNKEHFREHLDKLFAKGIGMLDIALNEAFNILSDFNHTGQGSICSQAIM
 LITDGAVDTYDTIFAKYNWPDRKVRIFTYILIGREAAFADNLKWMACANKGFFTQISTLADV
 QENVMEYLHVLSRPKVIDQEHDVVWTEAYIDSTLPQAQKLADDQGLVLMTTVAMPVFSKQN

ETRSKGILLGVGTDVPVKELLKTIPKYKLGIGHYAFAITNNGYILTHPELRPLYEEGKKR
 RKPNYSSVDLSEVEWEDRDDVLRNAMVNRKTGKFSMEVKKTVDKGKRVLVMNTDYYYTDIK
 GTPFSLGVALSRGHGKYFFRGNVITIEGLHDLEHPDVSLADEWSYCNTDLHPEHRHLSQLE
 AIKLYLKGKEPLLQCDKELIQEVLFDAVVSAPIEAYWTSALNKSSENSDKGVEVAFLGTRT
 5 GLSRINLFVGAEQLTNQDFLKAGDKENIFNADHFPLWYRRAAEQIAGSFVYSIPFSTGTVN
 KSNVVTASTSIQLLDERKSPVVAAGVIQMKLEFFQRKFWTASRQCASLDGKCSISCDETV
 NCYLIDNNGFILVSEDYTQTGDFFGVEGAVMNKLLTMGSFKRITLYDYQAMCRANKESSD
 SAHGLLDPYKAFLSAAKWIMTELVLFLVEFNLCSWHSDMTAKAQKLKQTLEPCDTEYPAF
 VSEKTIKETTGNIACEDCSKSFVIQQIPSSNLFMVVVDSSCLCESVAPITMAPIEIRYNES
 10 LKCERLKAQKIRRRPESCHGFHPEENARECGGASHHHHHH

25 - nucleotide sequence

TCGCCACCATGGCGGTGCCGGCTC

15 **26 - nucleotide sequence**

TCGGAATTCCTCAGTGATGGTGATGGTGATGGGCCCCGCGGCCACAGTC

27 - nucleotide sequence

TCGCCACCATGGCCGGGCGGGC

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28 - nucleotide sequence

TCTCAGTGATGGTGATGGTGATGCGATGCACCCCCACACTCTC

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CLAIMS

- 1) A purified or isolated nucleic acid encoding a mammalian secreted soluble cerebral cortical voltage-dependent calcium channel $\alpha_2\delta$ -2, $\alpha_2\delta$ -3 or $\alpha_2\delta$ -4 subunit polypeptide.
- 2) A purified or isolated nucleic acid according to claim 1, comprising a polynucleotide having at least 90% identity with the sequence encoding :
 - from amino-acid 1 to between amino-acids 1027 and 1062 of SEQ ID N°20 for $\alpha_2\delta$ -2,
 - from amino-acid 1 to between amino-acids 984 and 1019 of SEQ ID N°22 for $\alpha_2\delta$ -3.
- 3) A purified or isolated nucleic acid according to claim 1, having at least 90% identity with the sequence encoding :
 - from amino-acid 1 to between amino-acids 1047 and 1062 of SEQ ID N°20 for $\alpha_2\delta$ -2,
 - from amino-acid 1 to between amino-acids 1004 and 1019 of SEQ ID N°22 for $\alpha_2\delta$ -3.
- 4) A purified or isolated nucleotide sequence according to claim 1 wherein said sequence is the sequence of SEQ ID N°1, SEQ ID N°2, SEQ ID N°3, SEQ ID N°7, SEQ ID N°8, SEQ ID N°9, SEQ ID N°13, SEQ ID N°14, SEQ ID N°15, SEQ ID N°19 or SEQ ID N°21.
- 5) A purified or isolated nucleic acid, having at least 90% identity with the nucleotide sequence of SEQ ID N°19 or SEQ ID N°21.
- 6) A purified or isolated polynucleotide comprising at least 10 consecutive nucleotides of the nucleotide sequence of SEQ ID N°19 or SEQ ID N°21.
- 7) A polynucleotide probe or primer hybridizing, under stringent conditions, with the with the nucleotide sequence of SEQ ID N°19 or SEQ ID N°21.
- 8) A method for the amplification of a nucleic acid encoding a mammalian secreted soluble cerebral cortical voltage-dependent calcium channel $\alpha_2\delta$ -n subunit polypeptide wherein n is 2, 3 or 4, said method comprising the steps of:

(a) contacting a test sample suspected of containing the target secreted soluble $\alpha_2\delta$ -n subunit nucleic acid, or a sequence complementary thereto, with an amplification reaction reagent comprising a pair of amplification primers located on either side of the $\alpha_2\delta$ -n subunit nucleic acid region to be amplified, and

(b) optionally, detecting the amplification products.

9) A kit for the amplification of a nucleic acid encoding a secreted soluble $\alpha_2\delta$ -n subunit polypeptide wherein n is 2, 3 or 4, or a complementary sequence thereto in a test sample, wherein said kit comprises:

(a) a pair of oligonucleotide primers which can hybridize, under stringent conditions, to the secreted soluble $\alpha_2\delta$ -n subunit nucleic acid region to be amplified;

(b) optionally, the reagents necessary for performing the amplification reaction.

10) A recombinant vector comprising a nucleic acid according to claim 1.

11) A recombinant host cell comprising a nucleic acid according to claim 1.

12) A method for producing a secreted soluble $\alpha_2\delta$ -n subunit wherein n is 2, 3 or 4, and said method comprises the steps of:

(a) inserting the nucleic acid encoding the desired $\alpha_2\delta$ -n subunit polypeptide in an appropriate vector;

(b) culturing, in an appropriate culture medium, a host cell previously transformed or transfected with the recombinant vector of step (a);

(c) harvesting the culture medium thus obtained or lyse the host cell, for example by sonication or osmotic shock;

(d) separating or purifying, from said culture medium, or from the pellet of the resultant host cell lysate, the thus produced $\alpha_2\delta$ -n subunit polypeptide of interest.

13) A purified or isolated recombinant polypeptide comprising the amino acid sequence of a secreted soluble $\alpha_2\delta$ -2, $\alpha_2\delta$ -3 or $\alpha_2\delta$ -4 subunit polypeptide.

14) A recombinant polypeptide according to claim 14, having at least 80% amino-acid identity with a polypeptide comprising :

- from amino acid 1 to between amino acids 1027 and 1062 of the amino acid sequence of SEQ ID N°20, or
- from amino acid 1 to between amino acids 1019 and 1079 of the amino acid sequence of SEQ ID N°22.

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15) A recombinant polypeptide according to claim 14, wherein said recombinant polypeptide is selected from the group consisting of the amino acid sequences of SEQ ID n°4, SEQ ID n°5, SEQ ID n°6, SEQ ID n°10, SEQ ID n°11, SEQ ID n°12, SEQ ID n°16, SEQ ID n°17 and SEQ ID n°18.

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16) A method for the screening of ligands which bind a cerebral cortical voltage-dependent calcium channel $\alpha_2\delta$ -n subunit wherein n is 2, 3 or 4, said method comprising the steps of:

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- contacting a secreted soluble recombinant calcium channel $\alpha_2\delta$ -n subunit polypeptide with:

- a ligand of interest; and
- a labelled compound which binds the $\alpha_2\delta$ -n subunit; and
- measuring the level of binding of the labelled compound to the $\alpha_2\delta$ -n subunit.

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17) A method according to claim 16, wherein said method is a scintillation proximity assay.

18) A method according to claim 16, wherein said method is a flashplate assay.

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19) A method according to claim 16, wherein said method is a filter binding assay.

20) A method according to claim 16, wherein said secreted soluble recombinant calcium channel $\alpha_2\delta$ -n subunit polypeptide is selected from polypeptides having at least 80%, preferably 90%, more preferably 95%, and most preferably 98 or 99% amino-acid identity with the polypeptide comprising from amino acid 1 to between amino-acids 984 and 1063, preferably between amino-acids 994 and 1054, and most preferably between amino-acids 1019 and 1054 of SEQ ID N°5 or SEQ ID N°16.

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21) A method according to claim 16, wherein said secreted soluble recombinant calcium channel $\alpha_2\delta$ -n subunit polypeptide is selected from the group consisting of SEQ ID N°6, 7, 8, 9, 13, 14 and 15, with the polypeptides of SEQ ID N°9 and SEQ ID N°15 being most preferred.

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22) A kit for the screening of ligands which bind bind a cerebral cortical voltage-dependent calcium channel $\alpha_2\delta$ -n subunit wherein n is 2, 3 or 4, said kit comprising:

- a secreted soluble recombinant calcium channel $\alpha_2\delta$ -n subunit; and
- a labelled compound which binds to the $\alpha_2\delta$ -n subunit.

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ABSTRACT

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Secreted soluble $\alpha_2\delta$ -2, $\alpha_2\delta$ -3 or $\alpha_2\delta$ -4 calcium channel subunit polypeptides and screening assays using same

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Soluble $\alpha_2\delta$ sub-type polypeptides.

Methods for cloning, expression and purification of freely soluble $\alpha_2\delta$ subtype polypeptides.

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A method for the screening of ligands which bind to soluble $\alpha_2\delta$ subtype polypeptides.